

Research Article



Schistocephalus solidus (Cestoda: Ligulidae) Infection Characteristics and Pathology at the Highest Elevation Record Environment (Dalaman River Basin, Turkey)

SALİM SERKAN GÜÇLÜ¹, UFUĞ GÜRKAN YILDIRIM¹, DENİZ İNNAL², ÖZLEM ÖZMEN³

¹Isparta University of Applied Sciences, Eğirdir Fisheries Faculty, Isparta, Turkey; ²Burdur Mehmet Akif Ersoy University, Department of Biology, Burdur, Turkey; ³Burdur Mehmet Akif Ersoy University, Faculty of Veterinary Medicine, Department of Pathology, Burdur, Turkey.

Abstract | This study was aimed to identify *Schistocephalus solidus* in three-spined stickleback, *Gasterosteus aculeatus* (Teleostei: Gasterosteidae) and to describe infection parameters and pathology. Three-spined stickleback specimens were collected between October 2019 to September 2020 in Gökpınar Spring (Dalaman River basin-Turkey). The highest elevation record of the common cestod parasite of three-spined stickleback has been found in this environment. Gross examination revealed that abdominal swelling was the most common finding. Cachexia and growth retardation were common in infected fishes. During the necropsy parasites were localized at the abdominal cavity. At the histopathological examination of the parasite, marked inflammatory reaction and fibrous tissue proliferation were observed. This study result showed that *Schistocephalus solidus* can cause marked pathological findings in infected three-spined stickleback.

Keywords | Anatolia, high altitude, pathology, three-spined stickleback, Cestoda

Editor | Muhammad Imran Rashid, Department of Parasitology, University of Veterinary and Animal Sciences, Lahore, Pakistan.

Received | August 10, 2021; **Accepted** | September 14, 2021; **Published** | September 28, 2021

***Correspondence** | Salim Serkan GÜÇLÜ, Isparta University of Applied Sciences, Eğirdir Fisheries Faculty, Isparta, Turkey; **Email:** salimguclu@isparta.edu.tr

Citation | GÜÇLÜ SS, YILDIRIM UG, İNNAL D, ÖZMEN O (2021). *Schistocephalus solidus* (cestoda: ligulidae) infection characteristics and pathology at the highest elevation record environment (dalaman river basin, turkey). J. Adv. Parasitol. 8(3): 32-36.

DOI | <http://dx.doi.org/10.17582/journal.jap/2021/8.3.32.36>

ISSN | 2311-4096

Copyright © 2021 GÜÇLÜ et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

The Mediterranean region is a biodiversity hotspot for freshwater ecosystems, harboring many species (many of them endemic) and genetically distinct lineages that are of conservation concern (Araguas et al., 2012; Sharda et al., 2018). One of these species is the three-spined stickleback *Gasterosteus aculeatus*. It is a small teleost fish and is a model organism in evolutionary biology and ethology (Bell and Foster, 1994; Mäkinen et al., 2006; Cresko et al., 2007; Mäkinen and Merilä, 2008; Barber, 2013). It is distributed widely throughout the Holarctic region at latitudes ranging from 35° to 70° (Wootton, 1984). During the past two decades, the three-spined stickleback (*G. aculeatus*), a small teleost easily reared in laboratory

conditions, has become one of the most important model species in ecology and evolutionary biology (Fang et al., 2018). With a nearly circumpolar marine distribution in the Northern Hemisphere, this species has colonized freshwater habitats multiple times since the last glacial maximum, evolving dramatic morphological, physiological and behavioral adaptations in remarkably short periods of time (Barrett et al., 2011).

Sticklebacks are intermediate hosts to a number of parasites. Probably the most common, and certainly the most conspicuous of these is the large white, worm-like plerocercoid of the tapeworm *Schistocephalus solidus* (Maitland and Campbell, 1992). *Schistocephalus solidus*, a pseudophyllidean cestode, is threatening natural populations of *G. ac-*

uleatus. This tapeworm belongs to the Cestoda family of obligate parasitic tapeworms. Copepods and three-spined sticklebacks (*G. aculeatus*) are the first and second intermediate hosts, respectively, and piscivorous birds are the definitive hosts (Clarke, 1954; Dubinina, 1980). Sticklebacks become infected by eating the copepod crustaceans that themselves have been infected by ingesting the tapeworm eggs. These are released into the water from the faeces of a predatory bird, such as a gull, which is the final host-itself having been infected by swallowing an infected, probably disabled or dying, stickleback (Maitland and Campbell, 1992). The parasite has various effects on the physiology, phenotype, and behavior of its host (Hahn, 2020). The three-spined stickleback *S. solidus* host-parasite system has become an important model in experimental parasitology and is increasingly used to investigate a wide range of questions about host-parasite interactions and co-evolution (Barber and Scharsack, 2010).

Although plerocercoids of the *S. solidus* are common parasites of three-spined sticklebacks throughout the geographical range of the fish, there is no report from Turkey. There is little information regarding the parasitic infections of three-spined sticklebacks in different regions of Turkey (Özer, 2003; Öztürk and Özer, 2014; Öztürk and Özer, 2019). *Schistocephalus solidus* is a fairly common species; however, little is known about its altitudinal range. It lives in marine, brackish and inland water systems. The goal of this research is to add to our understanding of *Schistocephalus solidus* by reporting the species' highest elevation record. This paper's second goal is to give infection parameters. The third goal of this research is to assess the histological alterations that have occurred.

MATERIALS AND METHODS

The care of experimental animals was consistent with the Republic of Turkey animal welfare laws, guidelines and policies approved by Isparta University of Applied Sciences Local Ethics Committee for Animal Experiments (permit reference number 2020/001). The study was carried out in Gökpınar Spring (Dalaman River basin-Turkey) (37°20'37.3"N, 29°26'38.3"E - altitude: 850 m). Monthly samples were carried out in October 2019 and September 2020 with drift nets of tulle of 2 mm mesh size and 888 individuals were caught from Gökpınar Spring. Specimens of three-spined sticklebacks were placed in a well aerated 20 lt aquarium filled with stream water. The fish were maintained in the aquarium for 2-3 hours and subsequently anaesthetized by MS-222. The total length was measured and sex determined at necropsy by macroscopic investigation. During the dissection, abdominal cavity and visceral organs were examined separately under a dissecting microscope. Morphological identification of parasites

was performed following by Chubb et al. (1987). The parasite has been identified as *Schistocephalus solidus*.

Prevalence and mean intensity were calculated by Bush et al. (1997). During the dissection fish samples totally fixed in 10% neutral buffered formalin for 2 days. After fixation fish samples were placed to the tissue processing cassettes. Then samples were routinely processed by an automatic tissue processor (Leica ASP300S, Wetzlar, Germany). For tissue processing, samples dehydrated in ascending grades of ethanol, cleared in xylene, and embedded in paraffin wax. Afterward, serial sections of the paraffin blocks (5 µm thickness) were cut by using a rotary microtome (Leica RM 2155; Leica Microsystems, Wetzlar, Germany). Sections were stained with hematoxylin and eosin (HE) and examined using a light microscope. Microphotography was performed using the Database Manual Cell Sens Life Science Imaging Software System.

RESULTS

The total length of three-spined sticklebacks individuals varied between 1.38 and 7.08 cm, while the weight values ranged between 0.014 and 4.85 grams. Among the total number of 888 specimens of three-spined sticklebacks examined for parasite presence, 30 fish were infected with cestode *Schistocephalus solidus* (Figure 1, Table 1). The prevalence of infection was 3.38 % (sexes combined), while the mean intensity of infection during the study was 1.1 parasites per fish. The highest prevalence and mean intensity (5.83 % and 1.14 parasites/fish) was recorded in male host individuals (Table 1). Cestode infection was highest in summer (4.98%) and lowest in winter (1.41%) (Table 2). Abdominal swelling was the most common finding during the gross examination. Plerocercoids were found in free form in the abdominal cavity. Generally, a single plerocercoid was encountered, while more than one parasite was found in some fish. The very prominent tegument of the plerocercoid was elongated like a sawtooth from front to back at longitudinal sections (Figure 2). At the transversal sections the outer layer of the parasite was smooth. Microscopic protrusions were observed on the tegument. At least 4 circularly and longitudinally muscle layers were observed from inner to outer side of the plerocercoid. On the serosal surfaces of the organs of the fish that touched the parasite, vascular hyperemia and inflammatory cell infiltrations, mostly of mononuclear cells, were observed (Figure 3).

DISCUSSION

Due to the importance of *S. solidus* for zoogeographic distribution of three-spined sticklebacks, this paper presents a new record of a specific cestode parasite of three-spined sticklebacks in Turkey. *S. solidus* has been found at various

Table 1: Prevalence and Intensity of infection in sex groups of *G. aculeatus*

Sex	TL-cm (min-max)	W-g (min-max)	N	N'	P	Int.
Immature	2.78 (1.38-3.96)	0.26 (0.014-0.71)	265	3	1.13	1
Female	5.05 (2.25-7.08)	1.92 (0.14-4.85)	233	6	2.58	1
Male	4.17 (2.44-6.38)	0.96 (0.15-3.84)	360	21	5.83	1.14
Total	3.99 (1.38-7.08)	1.01 (0.014-4.85)	888	30	3.38	1.1

N = total number of hosts examined; N' = number of infected fishes; P = prevalence; Int = mean intensity of infection; TL= Total Length; W= Weight

Table 2: Seasonality of infection and mean infection intensity for *S. solidus* in *G. aculeatus* hosts

Season	N	N'	P	Total	Int.
Winter	142	2	1.41	2	1
Spring	172	6	3.49	6	1
Summer	261	13	4.98	16	1.23
Autumn	313	9	2.88	9	1
Total	888	30	3.38	33	1.1

N = total number of hosts examined; N' = number of infected fishes; P = prevalence; Int = mean intensity of infection

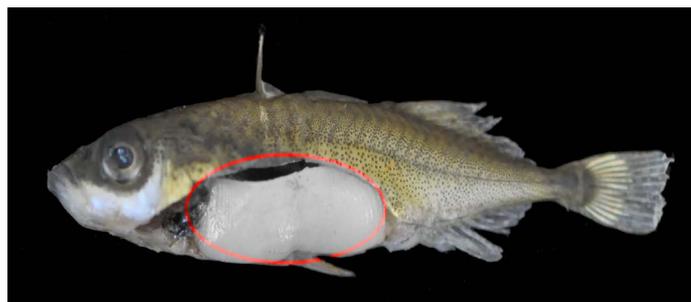


Figure 1: *G. aculeatus* individual infected with *Schistocephalus solidus*, 46.24 mm TL, May 2020 (*S. solidus*, circled in red.)

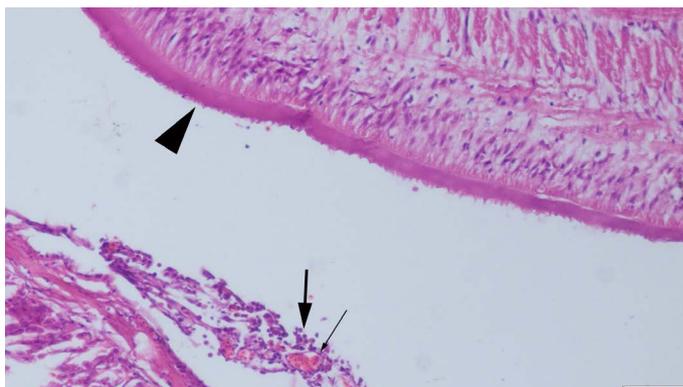


Figure 3: Serosal surface of the fish intestine, hyperemia of the vessels (thin arrow) and inflammatory cell infiltration (thick arrows) near the plerocercoid (arrow head), HE, Bar=50µm.

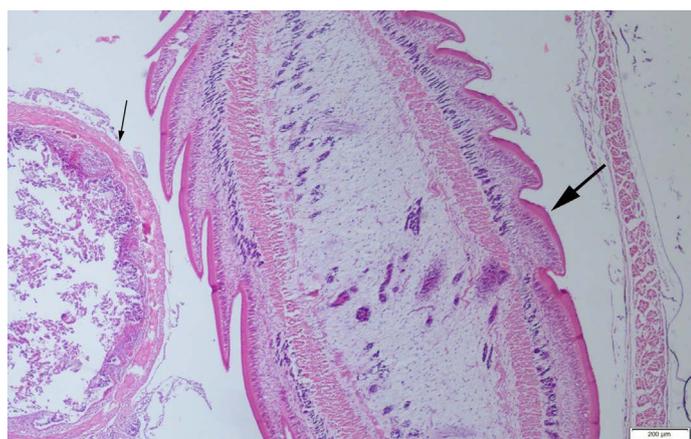


Figure 2: A plerocercoid (thick arrow) in abdominal cavity of the fish, near the intestine (thin arrow) of the host, HE, Bar=200µm.

elevations. The parasite was found at the highest altitude in the globe in the research location.

The prevalence of *S. solidus* in three-spined sticklebacks in this study (3.38 %) is lower than from other studies performed in Palearctic region. Bergersen (1996) found from 18% to 92% infected sticklebacks in freshwater localities in Greenland. Zander (2007) found a maximum of 14% sticklebacks infected with *S. solidus* in the Baltic Sea. Prevalence of *S. solidus* in the three-spined sticklebacks collected from a Gdynia Marina (Poland) was 94.4% at 2008 (Morozńska-Gogol, 2011). The reported prevalence of *S. solidus* was 54.2% from Puck Bay (Mačát et al., 2015). Different prevalence values in *S. solidus* infections between the

populations of three-spined sticklebacks may depend on the changing habitat conditions, physiology, immune parameters and health status of the fish. Different fish species infected with numerous parasites. Generally, hosts show no or little symptoms due to infection. Sometime clinical symptoms may occur at the tissue and organ level. Parasites can be localized all tissue or organ, but most common localization areas are abdominal cavity. Inflammatory reaction may occur if parasite remain in tissue for sufficient time. Common pathological findings were lymphocyte and macrophage infiltrations together with fibrous tissue proliferation around the parasite.

In this study, seasonal prevalence and histopathological findings in *G. aculeatus* caused by *S. solidus* was evaluated. The prevalence of the parasite was found higher in summer. At the histopathological examination all of the parasites were localized abdominal cavity. Parasites caused chronic inflammatory reaction characterized by mononuclear cell and macrophage infiltrations together with fibrous capsule around the parasites. This study revealed that *S. solidus* was the most common parasites of three-spined stickleback. This parasite caused clinical, gross and histopathological findings. The comparing the non-infected individuals infected ones were smaller and cachectic.

In fishes, parasitic infestations commonly occurs, prevalence of the infection may be higher in wild populations than cultured individuals (Roberts, 2012). Numerous parasites are reported in different fish species (Roberts, 2012). Pathology related the parasites, usually a combination of clinical signs, gross and histopathological findings of tissues and organs. Most of the parasites and reactions related the parasites can be identified during the hematoxylin and eosin stained sections (Bruno et al., 2006). Host responses against to parasites are important to evaluate parasite pathogenicity and it is generally assessed by histopathological examination (Yildiz et al., 2004). In addition, slight to marked abdominal swelling observed in infected fishes' gross examination, this study finding indicated that *S. solidus* caused marked inflammatory reaction in three-spined stickleback. But the inflammatory reaction was commonly composed mononuclear cells indicating the chronic inflammation. These findings showed that this parasite can persist for a long time without causing fish death.

Parasitic infections are generally chronic and slightly effect the host (Feist and Longshaw, 2005; Bourque et al., 2006). Acute parasitic infestations can cause high mortalities. Environmental factors such as temperature and dissolved oxygen concentration are the most important factors for parasitic infections (Feist and Longshaw, 2008). In this study, especially small number of parasites not caused the mortality according to gross and histopathological findings. Present study findings indicate that *S. solidus* may be

cause reduced condition or slower growing rate but it were not highly fatal parasitic infection for *Gasterosteus aculeatus*.

The habitat for the study is spring water. For this reason, the water temperature is almost constant. The high rate of infection during the summer months depends on day length rather than water temperature. Besides, the presence of kingfisher (*Alcedo atthis*) in the habitat environment has been determined. Webster et al. (2011) stated that kingfishers are the predators of *G. aculeatus*, while Raven (1986) stated that *G. aculeatus* constitute the majority of the food of kingfishers. These data also pose the risk of migration of the parasite to different habitats through birds.

In conclusion, the first record of *S. solidus* being seen in three-spined sticklebacks in Turkey is given in this study. It was also identified as an infected at the highest altitude.

CONFLICTS OF INTEREST

The authors have declared no conflict of interest.

AUTHORS CONTRIBUTION

The authors contributed equally.

REFERENCES

- Araguas R, Vidal O, Pla C, Sanz N (2012). High genetic diversity of the endangered Iberian three-spined stickleback (*Gasterosteus aculeatus*) at the Mediterranean edge of its range. *Freshwater Biol.* 57: 143–154. <https://doi.org/10.1111/j.1365-2427.2011.02705.x>
- Barber I (2013). Sticklebacks as model hosts in ecological and evolutionary parasitology. *Trends Parasitol.* 29: 556–566. <https://doi.org/10.1016/j.pt.2013.09.004>
- Barber I, Scharsack JP (2010). The three-spined stickleback - *Schistocephalus solidus* system: an experimental model for investigating host-parasite interactions in fish. *Parasitology*. 137(Special issue 3): 411–424
- Barrett RD, Paccard A, Healy TM, Bergesk S, Schulte PM, Schluter D, Rogers SM (2011). Rapid evolution of cold tolerance in stickleback. *Proceedings of the Royal Society B. Biol. Sci.* 278: 233–238. <https://doi.org/10.1098/rspb.2010.0923>
- Bell M, Foster S (1994). *The Evolutionary biology of the threespine stickleback*. Oxford University Press, New York.
- Bergersen R (1996). Sticklebacks from Greenland. *J. Fish Biol.* 48(4): 799–801. <https://doi.org/10.1111/j.1095-8649.1996.tb01474.x>
- Bourque JF, Dodson JJ, Ryan DAJ, Marcogliese DJ (2006). Cestode parasitism as a regulator of early life-history survival in an estuarine population of rainbow smelt *Osmerus mordax*. *Marine Ecology Progress Series*. 314: 295–307. <https://doi.org/10.3354/meps314295>
- Bruno DW, Nowak B, Elliot DG (2006). *Guide to the identification of fish protozoan and metazoan parasites in*

- stained tissue sections. *Dis. Aquat. Organisms*. 70: 1–36. <https://doi.org/10.3354/dao070001>
- Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997). Parasitology meets ecology on its own terms: Margolis et al, Revisited. *J. Parasitol.* 83(4): 575–583. <https://doi.org/10.2307/3284227>
 - Chubb JC, Pool DW, Veltkamp CJ (1987). A key to the species of cestodes (tapeworms) parasitic in British and Irish freshwater fishes. *J. Fish Biol.* 31: 517–543. <https://doi.org/10.1111/j.1095-8649.1987.tb05256.x>
 - Clarke AS (1954). Studies on the life cycle of the pseudophyllidean cestode *Schistocephalus solidus*. *Proceedings of the Zoological Society of London*. 124: 257–302. <https://doi.org/10.1111/j.1469-7998.1954.tb07782.x>
 - Cresko W, McGuigan K, Phillips P, Postlethwait J (2007). Studies on threespine stickleback developmental evolution: progress and promise. *Genetica*. 129: 105–126. <https://doi.org/10.1007/s10709-006-0036-z>
 - Dubinina MN (1980). Tapeworms (Cestoda, Ligulidae) of the fauna of the USSR. Amerind Publishing, New Delhi, India, 320 pp.
 - Fanga B, Merilä J, Ribeiro F, Alexandrec CM, Momigliano P (2018). Worldwide phylogeny of three-spined sticklebacks. *Molecul. Phylogenet. Evol.* 127: 613–625. <https://doi.org/10.1016/j.ympev.2018.06.008>
 - Feist SW, Longshaw M (2005). Myxozoan diseases of fish and effects on host populations. *Acta Zoolog. Sinica*. 51: 758–760.
 - Feist SW, Longshaw M (2008). Histopathology of fish parasite infections—importance for populations. *J. Fish Biol.* 73: 2143–2160. <https://doi.org/10.1111/j.1095-8649.2008.02060.x>
 - Hahn M (2020). The role of microbes in the interaction between the cestode parasite *Schistocephalus solidus* and its threespine stickleback host. Doctor of Philosophy Thesis, Marine and Atmospheric Sciences, Stony Brook University, 165 p.
 - Mačát Z, Bednařík A, Rulík M (2015). The three-spined stickleback (*Gasterosteus aculeatus*) infection with *Schistocephalus solidus* in Hel marina (Puck Bay, Baltic Sea, Poland). *Oceanolog. Hydrobiolog. Studies*. 44(1): 11–17. <https://doi.org/10.1515/ohs-2015-0002>
 - Maitland PS, Campbell RN (1992). *Freshwater Fishes of the British Isles*. Harper Collins Publishers, Somerset, 368 pp.
 - Mäkinen H, Cano JM, Merilä J (2006). Genetic relationships among marine and freshwater populations of the European three-spined stickleback (*Gasterosteus aculeatus*) revealed by microsatellites. *Molecul. Ecol.* 15: 1519–1534. <https://doi.org/10.1111/j.1365-294X.2006.02871.x>
 - Mäkinen H, Merilä J (2008). Mitochondrial DNA phylogeography of the three-spined stickleback (*Gasterosteus aculeatus*) in Europe—Evidence for multiple glacial refugia. *Molecul. Phylogenet. Evol.* 46(1): 167–182. <https://doi.org/10.1016/j.ympev.2007.06.011>
 - Morozínska-Gogol J (2011). Changes in levels of infection with *Schistocephalus solidus* (Müller 1776) of the three-spined stickleback *Gasterosteus aculeatus* (Actinopterygii: Gasterosteidae) from the Gdynia Marina. *Oceanologia*. 53(1): 181–187. <https://doi.org/10.5697/oc.53-1.181>
 - Özer A (2003). The occurrence of *Trichodina domerguei* Wallengren, 1897 and *Trichodina tenuidens* Faure-Fremiet, 1944 on three spined stickleback, *Gasterosteus aculeatus* L., 1758 found in a brackish and freshwater environment. *Acta Protozoolog.* 42: 41–46.
 - Öztürk T, Özer A (2014). Monogenean Fish Parasites, Their Host Preferences and Seasonal Distributions in the Lower Kızılırmak Delta (Turkey). *Turkish J. Fisheries Aquat. Sci.* 14: 367–378.
 - Öztürk T, Özer A (2019). Digenean metacercariae parasitic in fishes in Sarikum Lagoon Lake, Sinop, Turkish Black Sea coast: species diversity, seasonal occurrence and histopathological effects. *Acta Zoolog. Bulgarica*. 71(3): 443–452.
 - Raven P (1986). The size of minnow prey in the diet of young Kingfishers *Alcedo atthis*. *Bird Study*. 33: 6–11. <https://doi.org/10.1080/00063658609476884>
 - Roberts RJ (2012). *Fish Pathology*. Wiley Blackwell, West Sussex, 592 pp. <https://doi.org/10.1002/9781118222942>
 - Sharda S, Argenti E, Lucek K (2018). On the status of threespine stickleback (*Gasterosteus aculeatus*, Linnaeus 1758) in Lake Bracciano, Italy. *Fishes*. 3(17): 1–6. <https://doi.org/10.3390/fishes3010017>
 - Webster MM, Atton N, Hart PJB, Ward AJB (2011). Habitat-Specific Morphological Variation among Threespine Sticklebacks (*Gasterosteus aculeatus*) within a Drainage Basin. *PLoS ONE*. 6(6): e21060. <https://doi.org/10.1371/journal.pone.0021060>
 - Wootton RJ (1976). *The biology of the sticklebacks*, London Acad. Press., 387 pp.
 - Wootton RJ (1984). *A functional biology of sticklebacks*. Croom Helm, London, 265 pp. <https://doi.org/10.1007/978-1-4615-8513-8>
 - Yildiz K, Kabackci N, Yarim M (2004). Pathological changes of tench intestines infected with *Pomphorhynchus laevis*. *Rev. Med. Vet.* 155: 71–73.
 - Zander CD (2007). Parasite diversity of sticklebacks from the Baltic Sea. *Parasitol. Res.* 100(2): 287–297. <https://doi.org/10.1007/s00436-006-0282-0>