



Prevalence of Methicillin-resistant Staphylococcal Mastitis in Ruminants

SAOUSSENE SMAALI*, FATIMA BOUKAZOULA, LAMIA SAADI, IMEN BRAHEM

Faculty of Exact Sciences and Natural and Life Sciences. Larbi Tebessi University –Tebessa, Algeria.

Abstract | The objectives of this work are to study the prevalence of staphylococcal mastitis, evaluate the resistance profile of these strains to antibiotics and the frequency of methicillin-resistant staphylococci in order to better define an effective therapeutic and preventive strategy. Milk samples from 41 females (cow, ewe and goat) were collected aseptically, and in the first step bacteriological tests were performed, including isolation and identification tests, to study the sensitivity of isolated staphylococci to nine antibiotics, using the antibiotic susceptibility reference method (agar dilution). The results were interpreted according to the criteria of Veterinary recommendations of the antibiotic susceptibility testing committee of the French microbiology society 2018. The results of bacteriological analyses of milk showed a significant prevalence of methicillin-resistant staphylococci mastitis (19.45%), including 17.85% of CSN and 1.2% for MRSA. The study of the antibiotic resistance profile of these isolated staphylococcus strains showed the high resistance of tetracycline with a frequency of 75% for CSP and 25% for CSN. This during vancomycin remains the most active antibiotic (100%) for both types of staphylococci.

Keywords | Algeria, MRSA-L, CSP, CSN, Livestock

Received | September 09, 2019; **Accepted** | January 10, 2020; **Published** | March 25, 2020

***Correspondence** | Saoussene Smaali, Department of Applied Biology, Faculty of Exact Sciences and Natural and Life Sciences. Larbi Tebessi University –Tebessa, Algeria; **Email:** saoussene.smaali@univ-tebessa.dz

Citation | Smaali S, Boukazoula F, Saadi L, Brahem I (2020). Prevalence of methicillin-resistant staphylococcal mastitis in ruminants. *Adv. Anim. Vet. Sci.* 8(4): 428-432.

DOI | <http://dx.doi.org/10.17582/journal.aavs/2020/8.4.428.432>

ISSN (Online) | 2307-8316; **ISSN (Print)** | 2309-3331

Copyright © 2020 Smaali *et al.* This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

The pathology of the mammary gland is one of the main diseases encountered in Algerian dairy farms (Bouزيد *et al.*, 2011). It is a multifactorial disease. The risk of its occurrence can be increased by chemical, physical or traumatic factors. Even after several control programs have been implemented, mastitis is still a high incidence disease worldwide, resulting in very significant economic losses due to a decrease in the quality and quantity of milk production, an increase in the cost of therapeutic protocols and veterinary services (Oviedo-Boyso *et al.*, 2006).

The large use of antibiotics is the most common strategy of control of mastitis especially during the dry period by intramammary administration of long acting antibiotics in tubes has played a major role in the development of bacterial antibiotic resistance. This resistance to ATB is mainly due to the high consumption of antibiotics for their

treatment but also for their prevention (Angoujard, 2015).

The emergence of multiresistant bacteria (BMR) will increasingly confront clinicians with real therapeutic impasses (Sanders *et al.*, 2011). In addition, the transmission of this type of bacteria from animals to humans is possible either through direct contact or via the food chain from food and animal products contaminated by these bacteria (Petinaki and Spiliopoulou, 2012).

Staphylococcus is the germ most involved in mastitis in Algerian farms. The study of the epidemiology of this species has evolved considerably over time, due to its importance in veterinary and human medicine. The increase in infectious episodes caused by this multidrug-resistant pathogen, particularly strains of methicillin-resistant *S. aureus* (MRSA) of animal origin, especially those associated with livestock (MRSA-L), is intriguing to the scientific community and health professionals. In particular, since

their zoonotic potential has been demonstrated (Petinaki and Spiliopoulou, 2012; Verkade and Kluytmans, 2013).

Despite the definite danger that these MRSA-L strains represent to human health, their existence is useful for science by serving as a study model to better understand host-pathogen interactions and to identify virulence factors and predisposing factors for the development of clinical infection. This improved knowledge is likely to open up new perspectives for treatment and control programmes in order to regain control over this formidable pathogen that causes harm in terms of profitability on farms and sometimes threatens people's lives.

It is in this context that our study, whose objectives are to study the prevalence of staphylococcal mastitis, evaluate the resistance profile of these strains to antibiotics and the frequency of staph methicillin resistance in order to better define an effective therapeutic and preventive strategy.

MATERIALS AND METHODS

The study was carried out on milk samples from 41 females (ewes, cows and goats) with mastitis. The isolation and purification of the strains was performed on blood agar at 37°C. Bacteria identification was done by conventional methods in quality control laboratory of the Tebessa Public Health Department. The selective medium used for gram+ bacteria (Staphylococcus and Streptococci) was the Chapman medium.

The catalase test was performed for the identification of staphylococci (Catalase positive: Staphylococci, Catalase negative: Streptococci). The coagulase test was used to identify *S. aureus* (coagulase positive) The sensitivity of the identified staphylococci to nine (9) antibiotics (Ampicillin (AM) 10µg, Oxacillin (OX) 1µg, Clindamycin (DA) 2µg, Vancomycin(VA) 30µg, Tetracycline (TE) 30µg, Erythromycin (E) 15µg, Cefotaxim (CTX) 30µg, Ofloxacin (OFX) 5µg) was studied using the standard antibiotic susceptibility method (dilution in agar medium). The discs used come from the Biomerieux laboratory (France). The results were interpreted according to the veterinary recommendations of the Antibiogram Committee of the French Society of Microbiology (AC-FSM VET, 2018).

RESULTS AND DISCUSSION

In 41 milk samples, 84 bacterial strains were isolated, 73.81% were gram positive and 26.19% were gram negative (Table 1) Studies by Kadja et al. (2013) showed that 69.9% were gram positive bacteria and 30.10% were gram negative bacteria. The prevalence of staphylococcal mastitis was 61.91%, of which 57.15% were SCN, and 4.76% were CSP.

Table 1: Distribution of isolated bacteria according to their groups.

Group of bacteria		Number	Percentage
Gram negatives		22	26.19%
Gram positive	Staphylococcus CSP (61.91)	4	4.76%
	CSN	48	57.15%
	Streptococci	8	9.52%
	Bacillus	2	2.38%
Total		84	100%

Staphylococci have been the main cause of milk contamination during production (milking machine, infected cow, man, incomplete cleaning of the milking machine) (Thieulon, 2005).

In 2014, Smaali provided a prevalence of 72.4% for staphylococcal mastitis in sheep. On the other hand, Bergonier et al. (2005) found *S. aureus* in 1 to 8% of isolations in France, Spain and Sardinia. On the other hand, Beheshli and its collaborators (2010) contributed a rate of 88.40% for staphylococci isolated from milk.

The results of the antibiotic resistance profile study are reported in Figures 1, 2 and 3.

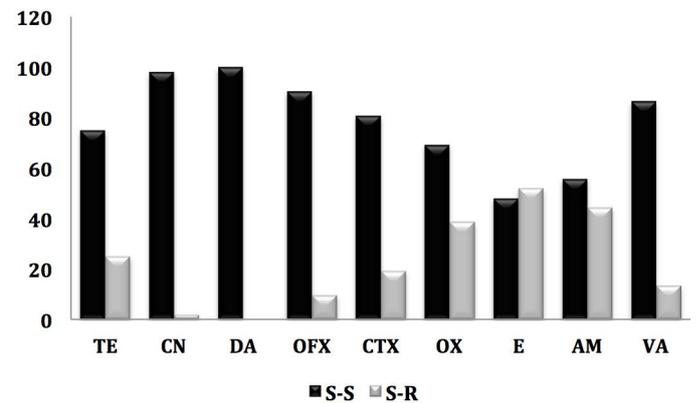


Figure 1: Antibiogram result of staphylococcus strains against 9 antibiotics. (S-S; e Staphylococcus-Sensitive, S-R; Staphylococcus-Resistant).

On all isolated (84 strains), the prevalence of methicilin-resistant staphylococci (oxicilin) was 19.45%, including 17.85% of CSN and 1.2% for MRSA. Indeed, work carried out in Saudi Arabia (Alzohairy, 2011) and Nigeria (Maisyama et al., 2014) reported prevalences of methecilin-resistant staphylococci by 15.5% and 28.8% respectively.

However, the detection of MRSA in sheep before slaughter once again confirms the risk to human health, not only for the consumer but also for all persons who come into contact with these animals (staff in establishments intended for the slaughter of animals, veterinarians, farmers) (Maisyama et al., 2014). Indeed, several studies have reported

that close and repeated contact with his animals is a risk factor for the acquisition of MRSA-L. In addition, some of these strains may cause serious infections (Van Cleef et al., 2011). These at-risk individuals can then serve as reservoirs for MRSA-L and ensure its deamination in the community (Lekkerkerk et al., 2015).

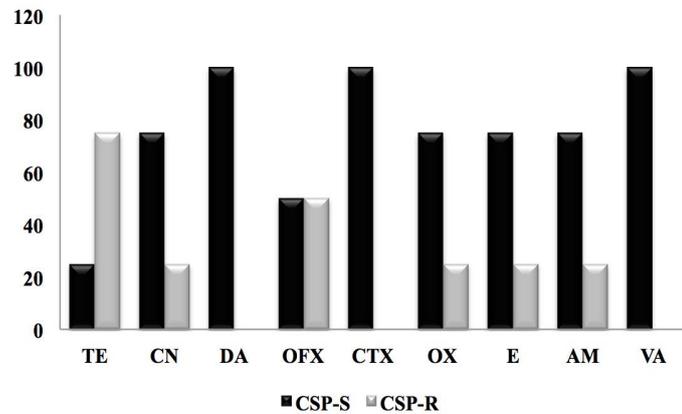


Figure 2: Antibiogram result of PCS strains against 9 antibiotics. (CSP-S; Coagulase Staphylococcus Positive-Sensitive, CSP-R; Coagulase Staphylococcus Positive-Resistant).

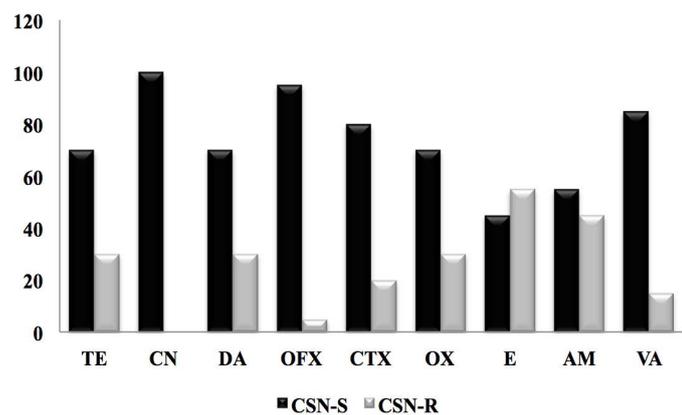


Figure 3: Result of the antibiotic susceptibility test of CSN strains against 9 antibiotics. (CSN-S; Coagulase Staphylococcus Negative-Sensitive, CSN-R; Coagulase Staphylococcus Negative-Resistant).

The results obtained (Figure 2) showed a very high resistance rate of CSP for tetracycline (75%), followed by ofloxacin (50%), oxacillin, (25%) erythromycin (25%) and ampicillin 25%. In contrast, CSP had 100% sensitivity to clindamycin, cefotaxime and vancomycin.

A recent study in the same region showed that SCPs were resistant to tetracycline (37.5%) and erythromycin (37.5%) and were sensitive to vancomycin (100%) (Smaali, 2019). Ben Hassen et al. (2003) showed that penicillin G resistance affected 64% of *S. aureus* strains, 36% for tetracycline and only 4% for oxacillin. For its part, Saidi (2014) provided a 100% resistance rate of *S. aureus* to Penicillin G.

In 2005, Turutoglu and al recorded a 61.1% resistance rate of *S. aureus* to Penicillin G in Turkey. Benhamed et al. (2011) isolated strains of *S. aureus* that were 100% methicillin sensitive.

In Malaysia, Jamali et al. (2014) obtained a resistance rate of 86% to penicillin and 76.6% to tetracycline.

The high percentage of resistance of *S. aureus* isolates to the b-lactamine family results from the acquisition of a B-lactamase gene and a plasmid penicillinase (Jamali et al., 2015). However, the resistance of these strains to Tetracycline is due to the extensive administration of this antibiotic on dairy farms (Jamali et al., 2015; Smaali, 2019).

For CSN strains, the antimicrobial resistance profile reported in Figure 3 showed a significant sensitivity of CSNs to clindamycin with a frequency (100%), followed by Ofloxacin (95%), vancomycin (85%) cefotaxime (80%), oxacillin and tetracycline with (70%), as well as ampicillin and erythromycin 55% and 45% respectively.

In 2003, Ben Hassen and al reported that CSNs have resistance to penicillin G with a percentage of (22.6%), tetracycline (20.8%), followed by streptomycin (16.5%) and erythromycin (16.5%), with low oxacillin resistance (1.7%).

Since their discovery, antibiotics have been used in veterinary medicine for curative purposes to treat sick animals, as a metaphysical treatment where animals in the same herd that are still healthy will receive the same treatment as sick animals and finally as a preventive or prophylactic measure to prevent certain diseases in animals identified as being at risk (Aarestrup, 2015). However, the misuse of antibiotics (insufficient doses, non-compliance with treatment duration) is most often at the root of this resistance.

The study of the antibiotic resistance profile of staphylococci (CSP and CSN) revealed the efficacy of vancomycin, clindamycin and ofloxacin against these pathogenic strains responsible for mastitis in runners.

CONCLUSION

The results of our study allowed us to conclude that the prevalence of methicillin-resistant staphylococcal mastitis was important (19.45%). The study of the resistance profile of the isolated strains shows the high resistance of these staphylococci to tetracycline. As well as the efficacy of vancomycin, clindamycin and ofloxacin against these pathogenic strains responsible for mastitis in runners. However, the presence of these multi resistant strains in ruminant livestock makes the antibiotic resistance problem in veterinary medicine even more complex than in human medicine.

To do this, Surveillance of these pathogenic bacteria in humans, animals and food allows us to identify the virulence factors of these pathogenic strains and establish appropriate control. However, the search for new, more effective molecules without side effects is necessary and we are moving towards the bioactive molecules of medicinal plants characteristic of Algeria, which will be the objective of our next articles.

In the meantime, it seems imperative to raise as much awareness as possible of the need to reduce the use of antibiotics as soon as possible, especially in veterinary medicine.

ACKNOWLEDGEMENT

The authors express their deep gratitude to Mr. R. Brakni and all the quality control laboratory staff (public health department-Tebessi. Algeria).

AUTHORS CONTRIBUTION

The research was designed jointly by Smaali Saoussene, Boukazoula Fatima, Saadi Lamia, Brahem Imen. Smaali Saoussene conducted the research. The authors read and approved the final manuscript.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- Aarestrup FM (2015). The livestock reservoir for antimicrobial resistance: a personal view on changing patterns of risks, effects of interventions and the way forward. *Phil. Trans. R. Soc. Lond. B. Boil. Sci.* 370(1670). <https://doi.org/10.1098/rstb.2014.0085>
- AC-FSM VET (2018). Veterinary recommendations of the antibiotic susceptibility testing committee of the French microbiology society. pp. 16. <https://doi.org/10.1111/jvp.12620>
- Alzohairy MA (2011). Colonization and antibiotic susceptibility pattern of methicillin resistance *Staphylococcus aureus* (MRSA) among farm animals in Saudi Arabia. *J. Bacteriol. Res.* 3: 63-68. <https://academicjournals.org/journal/JBR/article-full-text-pdf/6E59EF69690>
- Angoujard P (2015). Enquête sur le diagnostic et le traitement des mammites de la vache laitière par les vétérinaires de terrain en France. Thèse de doctorat Vétérinaire. École Nationale Vétérinaire D'Alfort. pp. 113. <http://theses.vet-alfort.fr/telecharger.php?id=1951>
- Beheshti R, Shaieghi J, Eshratkhan B, Jamshid GC, Nacer M (2010). Prévalence et étiologie des mammites subcliniques chez la brebis de la région de Tabriz, en Iran. *Vet. Glob.* 4 (3): 299-302. <https://www.idosijournals.org/gv/>

[gv4%283%2910/16.pdf](https://doi.org/10.21467/adv.v4i2.2833)

- Benhamed N, Moulay M, Aggad H, Henni JE and Kihal M (2011). Prevalence of mastitis infection and identification of causing bacteria in cattle in the oran region West Algeria. *J. Anim. Vet. Adv.*, 10(22): 3002-3005.
- Ben Hassen S, Messadi L, Ben Hassen A (2003). Identification et caractérisation des espèces de *Staphylococcus* isolées de lait de vaches atteintes ou non de mammites. *Ann. Méd. vét.* 147: 41-47. http://www.facmv.ulg.ac.be/amv/articles/2003_147_1_04.pdf
- Bergonier D, Lagriffouol G, Barillet F, Rupp R, Valognes A, Broujidoux R, Duquesnel R and Bertholot X (2005). Aetiological, clinical and epidemiological characterization of clinical mastitis in dairy sheep". Proceedings of the 4th IDF Int. Mastitis Conf., Maastricht, Netherlands, June 13-15, 2005 Wageningen Acad. Pub. Hogeveen edits, Wageningen, Netherlands. pp. 497-503.
- Bouzid R, Hocine A, Maifia F, Rezig F, Ouzrout R, Touati K (2011). Prévalence Des Mammites En Élevage Bovin Laitier Dans Le Nord-Est Algérien. *Livest. Res. Rural Dev.* 23(4): 73. <http://www.lrrd.org/lrrd23/4/bouz23073.htm>
- Chairat S, Gharsa H, Lozano C, Gomz-Sanz E, Gomez P, Zarazaga M, Boudabous A, Torres C, Ben Slama K (2015). Caractérisation of *Staphylococcus aureus* from Raw Meat Samples in Tunisia: detection of Clonal Lineage ST398 from the African Continent. *Foodborne path Dis.* 12: 686-692. <https://doi.org/10.1089/fpd.2015.1958>
- Jamali H, Radmehr B, Ismail S (2014). Short communication: prevalence and antibiotic Short communication: prevalence and antibiotic resistance of *Staphylococcus aureus* isolated from bovine clinical mastitis. *J. Dairy Sci.*, 97(4): 2226-2230. <https://doi.org/10.3168/jds.2013-7509>
- Jamali H, Paydar M, Radmehr B, Ismail S, Dadrasnia A (2015). Prevalence and antimicrobial resistance of *Staphylococcus aureus* isolated from raw milk and dairy products. *Food Control*, 54: 383-388. <https://doi.org/10.1016/j.foodcont.2015.02.013>
- Kadja M, Kane Y, Banah V, Kaboret Y (2013). Sensibilité Aux Antibiotiques Des Bactéries Associées Aux Mammites Cliniques Des Petits Ruminants Dans La Région De Dakar. *Ann. Sci. Agron.*, pp. 205-216. https://www.academia.edu/31480486/Sensibilit%C3%A9_Aux_Antibiotiques_Des_Bacteries_Associees_Aux_Mammites_Cliniques_Des_Petits_Ruminants_Dans_La_Region_De_Dakar
- Lekkerkerk WSN, van Wamel WJB, Snijders SV, Willems RJ, Van Duijkeren E, Broens EM, Wagenaar JA, Lindsay JA, Vosa MC (2015). What Is the Origin of Livestock-Associated Methicillin-Resistant *Staphylococcus aureus* clonal complex 398 isolates from humans without livestock contact? An epidemiological and genetic analysis. *J. Clin. Microbiol.* 53: 1836-1841. <https://doi.org/10.1128/JCM.02702-14>
- Mai-siyama IB, Okon KO, Adam NB, Askira UM, Isiaka TM, Adamu SJ, Mohammed A (2014). Methicillin resistant *Staphylococcus aureus* (MRSA) colonization rate among ruminant animals slaughtered for human consumption and contact person in Maiduguri, Nigeria. *Afr. J. Microbiol. Res.* 2643-2649. <https://doi.org/10.5897/AJMR2014.6855>
- Oviedo-Boyso, Valdez-Alarco N, Cajero-Jua Rez, Ochoa-Zarzosa Lopez-Meza Alejandro, Bravo-Patino, Baizabal-Aguirre (2006). Innate immune response of bovine mammary gland to pathogenic bacteria responsible for mastitis. *J. Infect.* 54: 399-409. <https://doi.org/10.1016/j.jinf.2006.06.010>

- Petinaki E, Spiliopoulou I (2012). Methicillin-resistant *Staphylococcus aureus* among companion and food-chain animals: impact of human contacts. Clin. Microbiol. Infect. 18: 626-634. <https://doi.org/10.1111/j.1469-0691.2012.03881.x>
- Saidi R, Khelef D, Kaidi R (2014). Bovine Mastitis: Prevalence of Bacterial Pathogens and Evaluation of Early Screening Test. Afr. J. Microbiol. Res., 7: 777-782.
- Sanders P, Bousquet-Melou A, Chauvin C, Toutain PL (2011). Utilisation des antibiotiques en élevage et enjeux de santé publique. <https://hal.archives-ouvertes.fr/hal-00939567>.
- Smaali S (2014). Etude de l'étiologie bactérienne des mammites subcliniques des ovins à l'Est de l'Algérie. Afrique Science 10(4): 225 – 231. <http://www.afriquescience.info/document.php?id=4105>
- Smaali S (2019). Antibiotic resistance profiles of staphylococci of animal and human origin. J. Exp. Appl. Anim. Sci. 3(1): 28-34. <https://doi.org/10.20454/jeaas.2019.1583>
- Thieulon M (2005). Laits pathogènes staphylocoques. Revue de la chambre d'agriculture du Cantal 12.
- Turutoglu H, Ercelik S and Ozoturk D (2006). Antibiotic resistance of *staphylococcus aureus* and coagulase-negative staphylococci isolated from bovine mastitis. Bull. Vet. Inst. Pulawy. 50: 41-45. <http://www.piwet.pulawy.pl/bulletin/images/stories/pdf/20061/20061041046.pdf>
- Van Cleef BA, Graveland H, Maenen AP, Van de Giessen AW, Heedrik D, Wadgenaar JA, Kluytmans JA (2011). Persistence of livestock-associated methicillin-resistant *Staphylococcus aureus* in field workers after short-term occupational exposure to pigs and veal calves. J. Clin. Microbiol. 49: 1030-1033. <https://doi.org/10.1128/JCM.00493-10>
- Verkade E, Kluytmans J (2013). Livestock-associated *Staphylococcus aureus* CC398: Animal reservoirs and human infections. Infect., Genet. Evol. 21: 523-530. <https://doi.org/10.1016/j.meegid.2013.02.013>