



# Development of Oocytes and Preimplantation Embryos of Mice Fed Diet Supplemented with *Dunaliella salina*

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**Abstract** | The present study investigated the developmental competence of germinal vesicle (GV) oocytes and the resulting embryos upon dietary *Dunaliella salina* (DS) supplementation. Quality of GV oocytes and maturation, timing of zygotes cleavage to two-cell stage, quality of embryos and offspring number and weight were investigated of mice supplemented with DS (100g/kg ration; N=30) compared to control (N=30). Females were injected with 7.5 I.U. of equine chorionic gonadotropin (PMSG) followed by 7.5 I.U. of human chorionic gonadotropin (hCG) after 48h and mated with fertile male. Immature GV oocytes were harvested from ovaries after 48h of PMSG injection for investigating quality of oocyte, timing of breakdown of germinal vesicle (GVBD) and extrusion of first polar bodies and maturation rate (%). Zygotes were harvested from uterine tubes 29–30h after hCG injection and followed for first cleavage whereas quality of blastocysts were evaluated after collection from uterine horn at 96h of hCG. Improvement of oocyte quality has been indicated in DS group whereas GVBD and maturation rate were not differed between DS and control groups. Although *D. salina* did not change timing of first cleavage of zygotes, blastocysts' quality were significantly ( $P < 0.05$ ) increased in DS group. Furthermore, offspring number ( $9.9 \pm 0.26$  &  $8.2 \pm 0.30$ ) and weight ( $12.01 \pm 0.31$  &  $9.84 \pm 0.37$ ) at birth were increased significantly ( $P < 0.05$ ) in the DS group compared to control one. In conclusion, supplementation of *D. salina* could increase reproductive performance through improvement quality of germinal vesicle oocytes and preimplantation embryos.

**Keywords** | Microalgae; *Dunaliella salina*; Oocytes, Embryos; Reproductive performances

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## INTRODUCTION

Recent interest has grown for the use of green microalgae including *Dunaliella salina* (*D. salina*) as a novel feed supplement for nutritional purposes (Yaakob et al., 2014) or detoxification and chelating of toxic and heavy metals. *D. salina* grant various health effects to the humans and animals as well. *D. salina* can protect against CNS oxygen toxicity (Bitterman et al., 1994) and gastrointestinal inflammation (Lavy et al., 2003) through anticancer and antioxidant properties because of its content of high  $\beta$ -carotene (Krinsky, 1988). Moreover, *D. salina* has immunomodulatory and anti-inflammatory effects (Hemmingson et al., 2006; Abdel-Daim et al., 2015). *D. salina* improved

body growth and reproduction (Nagasawa et al., 1989). Supplementation of algae or algal oil to dairy cattle increase n-3 percentage in milk fat without adverse effect on milk fat content (Stamey et al., 2012).

Stimulation of ovarian activity was widely explored through nutrition (Downing et al., 1995; Gifre et al., 2017), hormonal administration (Mandiki et al., 1993; Zarazaga et al., 1996), both nutritional and hormonal supplementation (Mohammed and Attaai, 2011). Because of the negative impacts of increased protein and energy in the diet on health and fertility (McEvoy et al., 1997; Dawuda et al., 2002) and the high cost of ration, it is important to search for non-traditional supplements with beneficial character-

istics for body. Microalgae possesses high concentration of proteins, lipids and vitamins, sulfated polysaccharides, several amounts of polyunsaturated fatty acids (Gouveia et al., 2008; Ghasemi et al., 2011; Wichuk et al., 2014; Cuelar-Bermudez et al., 2015; Gong and Bassi; 2016). Pulz and Gross (2004) showed that polysaccharides is important as therapeutic where it support immune system and has potential role to fight virus infection (Abdel-Daim et al., 2015). Several studies has been indicated that protein content in most of green microalgae is higher than plants (Becker, 1994; Becker, 2007). Microalgae consequently gift to humans and animals positive healthy and reproductive effects (Becker, 2007; Guedes et al., 2013; Choochote et al., 2014; Senosy et al., 2017).

The stimulatory effects of dietary green microalgae on reproductive performance has been confirmed. Conclusion of our work (Senosy et al., 2017) indicated that *D. salina* supplemented to Boer goats (10 g/day/head) accelerated ovarian follicle development and significantly increased follicle numbers (small, medium and large follicles). In addition, plasma metabolites and hormones (FSH, LH, estrogen and progesterone) were significantly increased upon supplementation of *D. salina*. Therefore, this study was conducted to explore beyond such information as development of oocytes and embryos. Kinetics of oocytes and embryos cleavages indicated differences either *in vitro* or *in vivo* (Mohammed et al., 2005; Mohammed and Attaai, 2011). First cleave of embryos and development was changed upon maturation of bovine oocytes in the presence of follicular fluid (FF) or fetal calf serum (FCS) (Mohammed et al., 2005). Because of the beneficial effects of *D. salina* on reproduction, the present work carried out to explore such improvement through the development of oocytes and embryos. Therefore, the objectives of the present research were to explore kinetics of oocyte and embryo cleavages in addition to reproductive performances upon feeding *D. salina* in mice.

## MATERIALS AND METHODS

All organic and inorganic materials used in this work were obtained from Caisson Lab (USA), unless stated otherwise. All media were prepared fresh and sterilized through 0.2- µm syringe filter (Nalgene, Cat. No. 190-2520).

### SITE OF STUDY AND MANAGEMENT

The study was conducted according to procedures approved by the Ethics Committee of Animal Experimentation of King Faisal University, Saudi Arabi. The study was conducted during the period from March to May 2017. Mice were kept controlled under cycle of 12h light and 12h dark. The controlled temperature and relative humidity during the experiment were 25 ±3°C and 50 ±10%, respective-

ly. Mice are fed commercial pellets of basal control diet (Arasco, KSA) or basal control diet supplemented with *D. salina* (100g/kg diet) and had free access to food and water for at least 4 wk.

### ANIMALS AND FEEDING

Sixty female albino mice of age 6-8 weeks were used during the study. The female were classified into two groups; the control group fed basal diet obtained from Arasco (KSA), which composed of 21.0%, 2.9% fat and 3.3% fiber, 1% mixture of vitamins and minerals (Table 1) and 3300 kcal/kg energy. In addition, *D. salina* algae added to basal diet (100 g/kg). *D. salina* microalgae were produced by Aquaculture National Group (KSA). The chemical composition of the experimental *D. salina* algae using standard methods (AOAC, 1990) was 8.47% ± 0.12moisture, 54.17% ± 0.47crude protein, 0.80% ± 0.10 fiber, 11.42% ± 0.22 total lipid, 18.47% ± 0.26 ether extract and 6.67% ± 0.23 ash.

**Table 1: The mixture of vitamins and minerals added to the basal control diet**

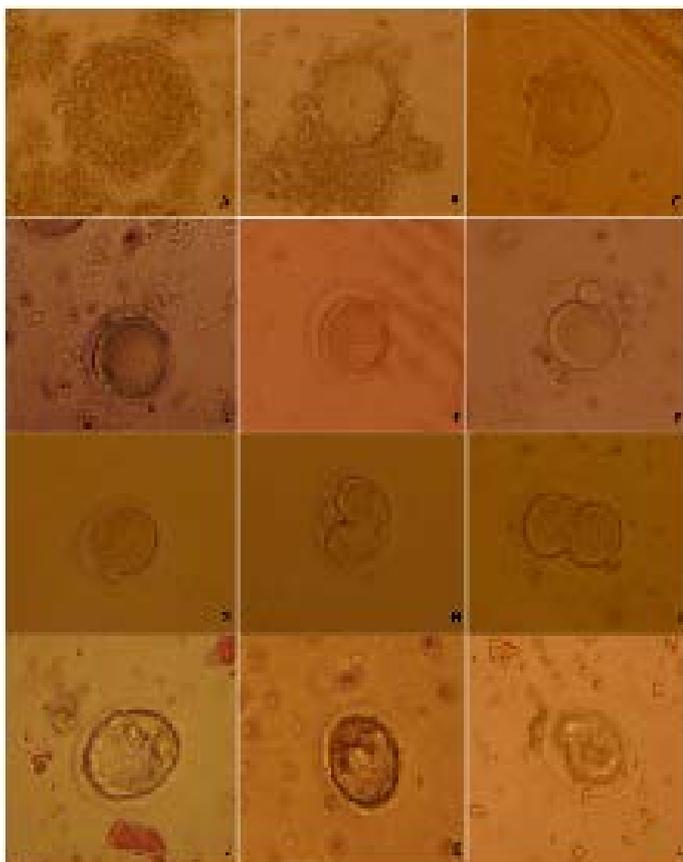
Vitamins mixture		Vitamins mixture	
Vitamin A	2000000, IU/kg	Folic acid	800, mg/kg
Vitamin D	300000, IU/kg	Pantothenic acid	5000, mg/kg
Vitamin E	40000, IU/kg	Biotin	200, mg/kg
Vitamin C	60000, mg/kg	Antioxidant	20000, mg/kg
Vitamin K	1200, mg/kg	Cobalt	200, mg/kg
Vitamin B1	4000, mg/kg	Copper	1000, mg/kg
Vitamin B2	4000, mg/kg	Iron	4000, mg/kg
Vitamin B6	2400, mg/kg	Manganese	15000, mg/kg
Vitamin B12	10, mg/kg	Selenium	20, mg/kg
Niacin	24000, mg/kg	Zinc	15000, mg/kg

### COLLECTIONS OF GERMINAL VESICLE OOCYTES

Females were given 7.5 I.U. of pregnant serum gonadotropin (PMSG, Intervet). Female mice were killed through cervical dislocation 44-48 h after PMSG injection. Ovaries were removed from the donor females. The GV-stage oocytes of ovarian follicles were released by puncturing with 30G sterile needles under a stereomicroscope. The cumulus-enclosed GV oocytes were released into HEPES tissue cell culture medium (TCM 199) supplemented with 5% fetal bovine serum. The cumulus-enclosed GV oocytes were immediately collected using glass pipette, its tip diameter larger than the diameter of cumulus-enclosed GV oocytes. The collected oocytes were graded into cumulus-enclosed GV oocytes (good oocytes), partial cumulus-enclosed GV oocytes (fair oocytes) and denuded oocytes (Figure 1A, B, C) (Mohammed and Attaai, 2011).

**TIMING OF GERMINAL VESICLE BREAKDOWN (GVBD) AND EXTRUSION OF FIRST POLAR BODY**

Cumulus cells were removed from oocytes using glass pipette, its tip diameter was smaller than cumulus-enclosed GV oocytes diameter. The denuded oocytes were cultured in TCM199 supplemented with 10.0% FCS. The oocytes were investigated for timing of GVBD and extrusion of first polar body during *in vitro* maturation (Figure 1D, E, F) (Mohammed et al., 2008; Mohammed et al., 2010; Mohammed, 2014).



**Figure 1:** Categorizing of developmental competence of oocytes and embryos upon *D. salina* supplementation in mice; A) whole cumulus-enclosed germinal vesicle (good) oocytes; B) partial cumulus-enclosed germinal vesicle (fair) oocytes; C) denuded (bad) oocytes ; D) germinal vesicle breakdown; E) processing of polar body extrusion stage; F) polar body extruded; G) late 1-cell stage embryo prior of cleavage; H) processing of first cleavage; I) two-cell stage embryo; J) Good quality embryo; K) fair quality embryo; L) bad quality embryo.

**COLLECTION OF CLEAVING EMBRYOS**

The females mice were super ovulated by 7.5 I.U. injection of equine chorionic gonadotropin (PMSG, Intervet) followed 48 h later by 7.5 I.U. injection of human chorionic gonadotropin (hCG, Intervet) and mated with fertile males. The same males were used to inseminate both females of control and DS group to exclude the paternal effect on embryo first cleavages. Late 1-cellstage embry-

os were collected in Dulbecco's modified medium supplemented with 10% FCS from the oviducts 29-30 h after hCG injection. The collected late 1-cell stage embryos were washed and cultured in KSOM+AA medium *in vitro* and followed for cleavage every 15 min (Figure 1 G, H, I).

**QUALITY OF COLLECTED EMBRYOS**

Embryos were flushed from the uteri at 4.00 days post hCG injection by using Dulbecco's modified medium supplemented with 10% FCS. The collected embryos graded into three categories; good, fair and bad embryos (Figure 1 J, K, L).

**REPRODUCTIVE PERFORMANCE**

Females were checked for parturition 4 times/day since day 18 of hCG injection. The numbers and weight of pups per female were counted and weighted for each group.

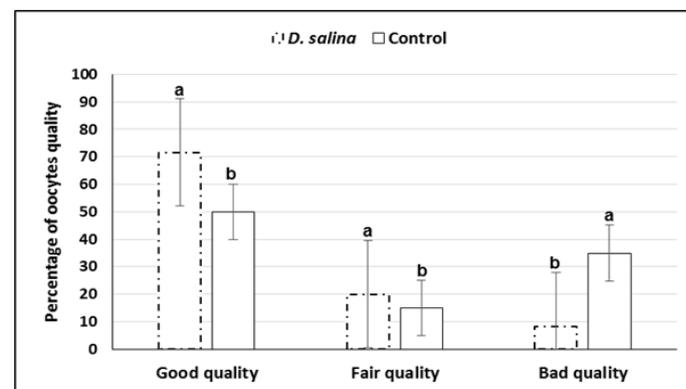
**STATISTICAL ANALYSIS**

Data are presented as mean ± SEM. The results were subjected to statistical analysis with independent t-test (SAS 2008). Comparison was done between control and *D. salina* treated groups. Values of probability of less than 0.05 (P < 0.05) were considered significant.

**RESULTS**

**OOCYTE QUALITY**

The GV-stage oocytes of ovarian follicles collected 44-48 h after PMSG injection were graded into good fair and bad quality. Quality of oocytes was improved upon *D. salina* supplementation. Percentage of cumulus enclosed GV (good) oocytes were significantly (P < 0.05) increased whereas denuded (bad) oocytes were significantly decreased in *D. salina* group compared to control one. The percentage of good, fair and denuded oocytes in *D. salina* group was 71.6± 1.29% (43/60), 20.0± 0.85% (12/60) and 8.4± 0.43% (5/60), respectively versus 50.0±1.29% (30/60), 15.0± 0.43 (9/60) and 35.0 ±0.85 (21/60) in control one (Figure 2).

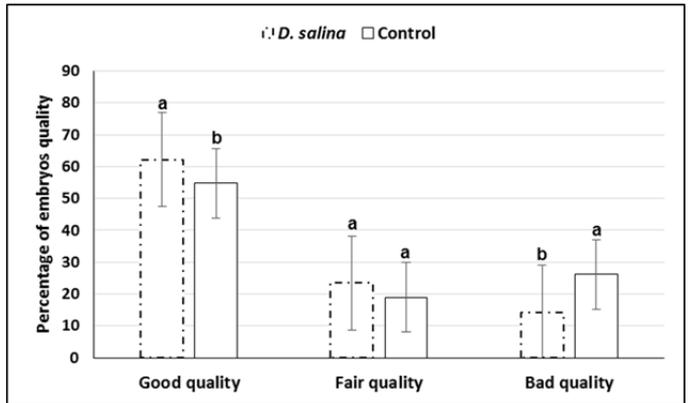


**Figure 2:** Quality of oocytes of *D. salina*(100g/kg diet) and control groups.

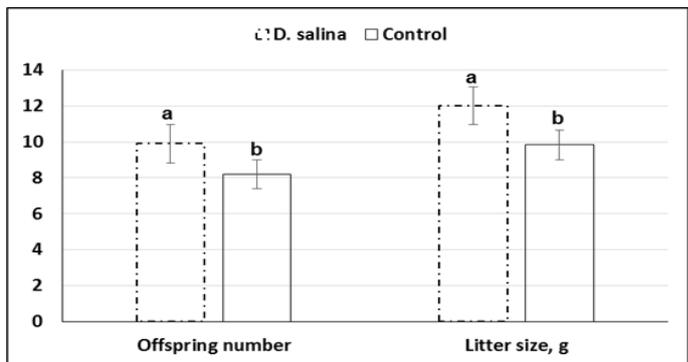
a,b; Values with different superscripts between *D. salina* and control groups significantly differed at P < 0.05.

**TIMING OF GERMINAL VESICLE BREAKDOWN (GVBD) AND EXTRUSION OF FIRST POLAR BODY**

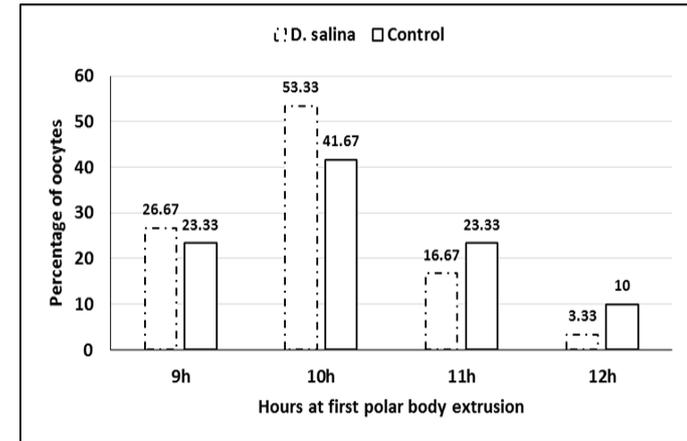
Sixty GV-stage oocytes cultured for *in vitro* maturation were investigated for timing of GVBD and extrusion of first polar body. No differences observed in timing of GVBD of *D. salina* and control oocytes, which occurred 2-3 hr. of starting maturation. Extrusion of 1<sup>st</sup> polar bodies of both *D. salina* and control oocytes occurred 9-12 h (Figure 3). It seems as *D. salina* supplementation slightly accelerated timing of polar body extrusion and did not change ( $P > 0.05$ ) percentage of oocyte maturation ( $96.66\% \pm 0.74; 58/60$  vs.  $95.0\% \pm 1.29; 57/60$ ).



**Figure 4:** Quality of embryos of *D. salina* (100g/kg diet) and control groups. a,b; Values with different superscripts between *D. salina* and control groups significantly differed at  $P < 0.05$ .



**Figure 5:** Offspring number and weight (g) of litter size of *D. salina* (100g/kg diet) and control groups. a,b; Values with different superscripts between *D. salina* and control groups significantly differed at  $P < 0.05$ .



**Figure 3:** Percentage of oocytes at timing extrusion of first polar body of *D. salina* and control oocytes.

**TIMING OF EMBRYO CLEAVAGE AND QUALITY AND REPRODUCTIVE PERFORMANCE**

Sixty late 1-cell stage embryos collected at 29-30 hr. of hCG injection of both *D. salina* and control were explored for first cleavages. Timing explored for first cleavage of late 1-cell stage embryos to two-cell stage embryos was not differed between *D. salina* and control group where all cleavages occurred within one hour and half. One hundred and thirty embryos were collected from ten females of *D. salina* group whereas one hundred and thirteen were collected from ten females of control group at day 4.0 of hCG injection. Percentage of embryos of good, fair and bad quality were significantly differed between *D. salina* and control groups. Percentage of embryos of good quality ( $62.16\% \pm 1.09$  vs.  $54.77\% \pm 1.38$ ) and fair quality ( $23.51\% \pm 2.46$  vs.  $18.97\% \pm 2.40$ ) were increased in *D. salina* group compared to control one. On the other hand, Percentage of embryos of bad quality were decreased in *D. salina* group compared to control one ( $14.30\% \pm 1.84$  vs.  $26.24\% \pm 1.72$ ) (Figure 4).

Furthermore, average of offspring number ( $9.9 \pm 0.26$  &  $8.2 \pm 0.30$ ) and weight ( $12.01 \pm 0.31$  &  $9.84 \pm 0.37$ ) at birth of ten deliveries were increased significantly ( $P < 0.05$ ) in the *D. salina* group compared to control one (Figure 5).

**DISCUSSION**

In the current study, the effects of *D. salina* supplementation on development of oocytes and embryos in addition to reproductive performance of mice were explored. *D. salina*, as an important type of green microalgae, supplementation improved oocyte and embryos development. Furthermore, reproductive performances including number and weight at birth were improved significantly ( $P < 0.05$ ) upon *D. salina* supplementation. Nutritional supply, *D. salina*, may be used as an alternative to hormonal treatments to enhance reproductive performance. *D. salina* has high levels of beta-carotene, glycerol, protein and other fine chemicals (Gouveia et al., 2008; Ghasemi et al., 2011; Wichuk et al., 2014; Cuellar-Bermudez et al., 2015; Gong and Bassi, 2016).

**OOCYTE QUALITY**

There are significant increases of cumulus-enclosed GV-oocytes ( $71.6\% \pm 1.29$  vs.  $50.0\% \pm 1.29$ ) and partial cumulus-enclosed GV-oocytes ( $20.0\% \pm 0.85$  vs.  $15.0\% \pm 0.43$ ), respectively in *D. salina* group compared to control

one. *D. salina* has high levels of beta-carotene, glycerol, protein and other fine chemicals (Gouveia et al., 2008; Ghasemi et al., 2011; Wichuk et al., 2014; Cuellar-Bermudez et al., 2015; Gong and Bassi, 2016), which might improve the quality of oocyte and support further embryonic development. Conclusion of our work (Senosy et al., 2017) indicated that *D. salina* supplemented to Boer goats (10 g/day/head) accelerated ovarian follicle development and significantly, increased follicle numbers (small, medium and large follicles). The improvement of oocyte quality in *D. salina* group compared to control one might be related to high levels of beta-carotene and elevation of plasma metabolites and hormones levels. Short-term beta-carotene supplementation in goat affects positively on serum insulin concentrations and ovarian activity (Meza-Herrera et al., 2013). Several studies suggest a direct role for insulin action on female reproduction. Senosy et al. (2017) found that *D. salina* supplementation to Boer goats resulted in significant increase of plasma FSH hormone levels, which rescued cumulus cell apoptosis and enhanced developmental capacity of buffalo oocytes (Jain et al., 2016).

#### TIMING OF GERMINAL VESICLE BREAKDOWN (GVBD) AND EXTRUSION OF FIRST POLAR BODY

No differences observed in timing of GVBD of *D. salina* and control oocytes, which occurred 2-3 hr. of starting maturation. Timing extrusion of 1<sup>st</sup> polar bodies of both *D. salina* and control oocytes occurred 9-12 hr. It seems as *D. salina* supplementation slightly accelerated timing of polar body extrusion and did not change percentage of oocyte maturation ( $96.66\% \pm 0.74$  vs.  $95.0\% \pm 1.29$ ). Timing of GVBD and extrusion of first polar body affects arrangements of chromosomes during maturation and further embryonic development (Mohammed et al., 2008; Mohammed et al., 2010; Mohammed, 2014). It seems that the timing occurred of germinal vesicle breakdown and polar body extrusion supported further embryonic development and reproductive performance.

#### TIMING OF FIRST EMBRYO CLEAVAGE AND EMBRYO QUALITY AND REPRODUCTIVE PERFORMANCE

Although timing explored for first cleavage of late 1-cell stage embryos to two-cell stage embryos was not differed between *D. salina* and control group, percentage of embryos of good quality ( $62.16\% \pm 1.09$  vs.  $54.77\% \pm 1.38$ ;  $P < 0.05$ ) and fair quality ( $23.51\% \pm 2.46$  vs.  $18.97\% \pm 2.40$ ;  $P > 0.05$ ) were increased in *D. salina* group compared to control one. The improvement of developmental capacity of oocytes and embryos due to supplementation of female mice with *D. salina* (100g/kg diet) confirmed by the significant ( $P < 0.05$ ) increase of number of litter size ( $9.9 \pm 0.26$  Vs.  $8.2 \pm 0.30$ ;  $P < 0.023$ ) and weight ( $12.02 \pm 0.31$  Vs.  $9.84 \pm 0.37$ ;  $P < 0.023$ ) at birth compared to control group. The improvement of number of litter size and weight might be attributed also to the beneficial effects of

*D. salina* on body health and ovarian function. The recorded body weight during the experimental period indicates increasing of body weight gain by about 10.0% of *D. salina* group compared to control one as in other studies (Nagasawa et al., 1989).

High concentrations of essential fatty acids (EFAs) found in *D. salina* algae. These EFAs (Omega 3, Omega 6, linoleic acid) support function of normal reproduction and carotenoids absorption, vitamin E and other fat-soluble nutrients. There are beneficial effects of beta-carotene of *D. salina* on reproduction reported of various species. beta-carotene administration in mice (Chew and Arche, 1983) and pigs (Brief and Chew, 1985; Kostoglou et al., 2000) increased litter size and decreased embryonic mortality. Haliloglu et al. (2002) found a positive correlation between beta-carotene level and corpus luteum weight and function of pregnant cattle. In addition, dietary beta-carotene supplementation in dairy cows decreased endometritis and retained placental (Michal et al., 1994). Schweigert et al. (2002) studied the effect of dietary beta-carotene of gilts on uterine fluid composition and early embryonic development. They found that beta-carotene resulted in a higher number of early developing embryos. beta-carotene injections to super ovulated Holstein cows improved embryo quality (Sales et al., 2008).

There are significant increases upon *D. salina* supplementation of blood and plasma metabolites and hormones (FSH, LH, estrogen and progesterone) concentrations indicated in several studies (Alishahi et al., 2014; Senosy et al., 2017). Such improvement of follicles development in addition to the increase of plasma metabolites and hormones (Senosy et al., 2017), which positively effect on the subsequent quality of oocytes and embryos might be the reason of the significant ( $P < 0.05$ ) increase of number and weight of litter size at birth in this study.

#### CONCLUSION

Supplementation of *D. salina* could increase reproductive performance of mice through the improvement of oocyte quality and preimplantation embryo development. Further *in vitro* studies are still required to investigate the developmental competence of oocytes and embryos upon *D. salina* supplementation.

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#### CONFLICT OF INTEREST

No conflict of interest for author to declare.

All authors contributed equally.

## REFERENCES

- Abdel-Daim MM, Farouk SM, Madkour FF, Azab SS (2015). Anti-inflammatory and immunomodulatory effects of *Spirulina platensis* in comparison to *Dunaliella salina* in acetic acid-induced rat experimental colitis. *Immunopharmacol. Immunotoxicol.* 37 (2):126- 139. <https://doi.org/10.3109/08923973.2014.998368>
- Alishahi M, Karamifar M, Mesbah M, Zarei M (2014). Hemato-immunological responses of *Heros severus* fed diets supplemented with different levels of *Dunaliella salina*. *Fish Physiol. Biochem.* 40: 57–65. <https://doi.org/10.1007/s10695-013-9823-5>
- AOAC (1990). Official methods of analysis of the AOAC. 15<sup>th</sup> Ed. [Arlington, VA, USA].
- Becker EW (1994). *Microalgae Biotechnology and Microbiology*. Cambridge University, New York, USA.
- Becker EW (2007). Micro-algae as a source of protein. *Biotechnol. Adv.* 25: 207-210. <https://doi.org/10.1016/j.biotechadv.2006.11.002>
- Bitterman N, Melamed Y, Ben-Amotz A (1994). b-Carotene and CNS oxygen toxicity in rats. *J. Appl. Physiol.* 76:1073–1076. <https://doi.org/10.1152/jappl.1994.76.3.1073>
- Brief S, Chew BP (1985). Effects of vitamin A and beta-carotene on reproductive performance in gilts. *J. Anim. Sci.* 60: 998–1004. <https://doi.org/10.2527/jas1985.604998x>
- Chew BP, Arche RG (1983). Comparative role of vitamin A and beta-carotene on reproduction and neonate survival in rats. *Theriogenol.* 20: 459–472. [https://doi.org/10.1016/0093-691X\(83\)90205-4](https://doi.org/10.1016/0093-691X(83)90205-4)
- Choochote W, Suklampoo L, Ochaikul D (2014). Evaluation of antioxidant capacities of green microalgae. *J. Appl. Phycol.* 26 (1): 43–48. <https://doi.org/10.1007/s10811-013-0084-6>
- Cuellar-Bermudez SP, Aguilar-Hernandez I, Cardenas-Chavez DL, Ornelas-Soto N, Romero-Ogawa MA, Parra-Saldivar R (2015). Extraction and purification of high value metabolites from microalgae: essential lipids, astaxanthin and phycobiliproteins. *Microb. Biotechnol.* 8: 190–209. <https://doi.org/10.1111/1751-7915.12167>
- Dawuda PM., Scaramuzzi RJ, Leese HJ, Hall CJ, Peters AR, Drew SB, Wathes DC (2002). Effect of timing of urea on the yield and quality of embryos in lactating dairy cows. *Theriogenol.* 58: 1443–1455. [https://doi.org/10.1016/S0093-691X\(02\)00973-1](https://doi.org/10.1016/S0093-691X(02)00973-1)
- Downing JA, Joss J, Scaramuzzi RJ (1995). Ovulation rate and the concentrations of gonadotrophins and metabolic hormones in ewes infused with glucose during the late luteal phase of the oestrous cycle. *J. Endocrinol.* 146: 403–410. <https://doi.org/10.1677/joe.0.1460403>
- Ghasemi Y, Rasoul-Amini S, Morowvat MH (2011). Algae for the production of SCP. In: Liang, M.T. (Ed.), *Bioprocess Sciences and Technology*. Nova Science Publishers Inc. 163–184.
- Gifre L, Arís A, Bach À, Garcia-Fruitós E (2017). Trends in recombinant protein use in animal production. *Microb. Cell Fact.* 16: 40. <https://doi.org/10.1186/s12934-017-0654-4>
- Gong M, Bassi A (2016). Carotenoids from microalgae: A review of recent developments. *Biotech. Adv.* 34: 1396–1412. <https://doi.org/10.1016/j.biotechadv.2016.10.005>
- Gouveia L, Batista AP, Sousa I, Raymundo A, Bandarra NM (2008). Microalgae in novel food products. In K. Papadopoulos, *Food Chemistry Research Developments*, pp. 75–112. New York: Nova Science Publishers.
- Guedes AC, Gião MS, Seabra R, Ferreira ACS, Tamagnini P, Moradas-Ferreira P, Malcata FX (2013). Evaluation of the antioxidant activity of cell extracts from microalgae. *Mar. Drugs.* 11 (4): 1256–1270. <https://doi.org/10.3390/md11041256>
- Haliloglu S, Baspinar N, Serpek B, Erdem H, Bulut Z (2002). Vitamin A and beta-carotene levels in plasma, corpus luteum and follicular fluid of cyclic and pregnant cattle. *Reprod. Dom. Anim.* 37(2):96–99. <https://doi.org/10.1046/j.1439-0531.2002.00338.x>
- Hemmingson JA, Falshaw R, Furneaux RH, Thompson K (2006). Structure and antiviral activity of the galactofucan sulfates extracts from *Undaria pinnatifida* (Phaeophyta). *J. Appl. Phycol.* 18(2):185-193. <https://doi.org/10.1007/s10>
- Jain A, Jain T, Kumar P, Kumar M, De S, Gohain M, Kumar R, Datta TK (2016). Follicle-stimulating hormone-induced rescue of cumulus cell apoptosis and enhanced development ability of buffalo oocytes. *Domest. Anim. Endocrin.* 55: 74–82. <https://doi.org/10.1016/j.domaniend.2015.10.007>
- Kostoglou P, Kyriakis SC, Papasteriadis A, Roumpies N, Alexopoulos C, Saoulidis K (2000). Effect of beta-carotene on health status and performance of sows and their litters. *J. Anim. Physiol. Anim. Nutr.* 83: 150–157. <https://doi.org/10.1046/j.1439-0396.2000.00263.x>
- Krinsky NI (1988). Membrane antioxidants. *Ann NY Acad. Sci.* 551: 17-32. <https://doi.org/10.1111/j.1749-6632.1988.tb22317.x>
- Lavy A, Naveh Y, Coleman R, Mokady S, Werman MJ (2003). Dietary *Dunaliella bardawil*, a beta-carotene-rich alga, protects against acetic acid-induced small bowel inflammation in rats. *Inflamm. Bowel. Dis.* 9(6):372-379. <https://doi.org/10.1097/00054725-200311000-00005>
- Mandiki SN, Bister JL, Paquay R (1993). Effects of hormonal manipulation on the resumption postpartum reproductive activity in Texel ewes. *Theriogenol.* 40: 607-620. [https://doi.org/10.1016/0093-691X\(93\)90413-Y](https://doi.org/10.1016/0093-691X(93)90413-Y)
- McEvoy TG, Robinson JJ, Aitken RP, Findlay PA, Robertson IS (1997). Dietary excesses of urea influence the viability and metabolism of preimplantation sheep embryos and may affect fetal growth among survivors. *Anim. Reprod. Sci.* 47(1-2):71-90. [https://doi.org/10.1016/S0378-4320\(96\)01627-2](https://doi.org/10.1016/S0378-4320(96)01627-2)
- Meza-Herrera CA, Vargas-Beltran F, Tena-Sempere M, González-Bulnes A, Macias-Cruz U, Veliz-Deras FG (2013). Short-term beta-carotene-supplementation positively affects ovarian activity and serum insulin concentrations in a goat model. *J. Endocrinol. Invest.* 36(3): 185-189.
- Michal JJ, Heirman LR, Wong TW, Chew BP, Frigg M, Volker L (1994). Modulatory effects of dietary beta-carotene on blood and mammary leukocyte function in periparturient dairy cows. *J. Anim. Sci.* 77: 1408– 1421.
- Mohammed AA (2014). Is nucleo-cytoplasmic incompatibility the reason of acceleration polar body extrusion? *Int. J. Curr. Eng. Sci. Res.* 4 (1): 287-291.
- Mohammed AA, Attaai AH (2011). Effects of dietary urea on timing of embryo cleavages and blood components in mice. *Vet. World.* 4: 360-363.
- Mohammed AA, j Karasiewicz, J Kubacka, P Greda, JA Modlinski (2010). Enucleated GV oocytes as recipients of

- embryonic nuclei in the G1, S, or G2 stages of the cell cycle. *Cell. Reprogram.* 12 (4): 427-435. <https://doi.org/10.1089/cell.2009.0107>
- Mohammed AA, J Karasiewicz, JA Modlinski, K Papis (2005). Oocyte maturation in the presence of randomly pooled follicular fluid increases bovine blastocyst yield in vitro. *J. Anim. Feed Sci.* 14: 501-512. <https://doi.org/10.22358/jafs/67048/2005>
  - Mohammed AA, Karasiewicz J, Modliński JA (2008). Developmental potential of selectively enucleated immature mouse oocytes upon nuclear transfer. *Mol. Reprod. Dev.* 75 (12): 1269-1280. <https://doi.org/10.1002/mrd.20870>
  - Nagasawa H, Konishi R, Yamamoto K, Ben-Amotz A (1989). Effects of beta-carotene-rich algae *Dunaliella* on reproduction and body growth in mice. *In Vivo.* 3(2):79-81.
  - Pulz O, Gross W (2004). Valuable products from biotechnology of microalgae. *Appl. Microbiol. Biotechnol.* 65(6): 635-648 <https://doi.org/10.1007/s00253-004-1647-x>.
  - Sales JNS, Dias LM, Viveiros AT, Pereira MN, Souza JC (2008). Embryo production and quality of Holstein heifers and cows supplemented with beta-carotene and tocopherol. *Anim. Reprod. Sci.* 106: 77-89. <https://doi.org/10.1016/j.anireprosci.2007.04.001>
  - SAS (2008). Statistical Analysis System. SAS statistics. Guide release, SAS Institute Inc., Cary, NC, USA.
  - Schweigert FJ, Krieger K, Schnurrbusch U, Schams D, Gropp J (2002). Effect of dietary beta-carotene on the early embryonic development and uterine fluid composition of gilts. *J. Anim. Physiol. Anim. Nutr. (Berl).* 86(7-8): 265-72. <https://doi.org/10.1046/j.1439-0396.2002.00384.x>
  - Senosy W, AY Kassab, AA Mohammed (2017). Effects of feeding green microalgae on ovarian activity, reproductive hormones and metabolic parameters of Boer goats in arid subtropics. *Theriogenol.* 96: 16-22. <https://doi.org/10.1016/j.theriogenology.2017.03.019>
  - Stamey JA, Shepherd DM, de Veth MJ, Corl BA (2012). Use of algae or algal oil rich in n-3 fatty acids as a feed supplement for dairy cattle. *J. Dairy Sci.* 95 5269-5275. <https://doi.org/10.3168/jds.2012-5412>
  - Wichuk K, Brynjólfsson S, Fu W (2014). Biotechnological production of value-added carotenoids from microalgae. *Bioengineered.* 95(9): 5269-5275. <https://doi.org/10.4161/bioe.28720>
  - Yaakob Z, Ali E, Zainal A, Mohamad M, Takriff MS (1996). An overview: biomolecules from microalgae for animal feed and aquaculture. *J Biol Res (Thessalon).* 21(1): 6. <https://doi.org/10.1186/2241-5793-21-6>
  - Zarazaga LA, Malpoux B, Chemineau P (1996). Characteristics of the plasma melatonin rhythm are not modified by steroids during the estrous cycle in Ile-de-France ewes. *J. Pineal Res.* 21: 114-120. <https://doi.org/10.1111/j.1600-079X.1996.tb00278.x>