Case Report

Therapeutic Management of Clinical Bovine Johne’s disease Using Goat Based ‘Indigenous Vaccine’ in Native Hariana Cattle: Case Reports

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ABSTRACT

Study reports therapeutic management of clinical to advance clinical Bovine Johne’s disease in a young male calf and an adult dairy cow of native Hariana breed in Mathura district. Calf (14 months old) suffering from symptoms of clinical bovine Johne’s disease (weakness and stunting without diarrhoea) was negative for MAP infection in fecal microscopy, blood PCR and ELISA. An adult cow (144 months old) a case of advance clinical BJD (suffering from non-treatable diarrhoea for last 3 years and weakness) was positive in ELISA and negative in microscopy and blood PCR tests. After exhausting all possible therapies, both the animals were vaccinated with 2 ml of ‘indigenous vaccine’ made from ‘Indian Bison Type’ biotype of Mycobacterium avium subspecies paratuberculosis (strain S 5) of goat origin, as last option. ‘Indigenous vaccine’ effectively helped in the management of clinical JD in case of calf however immune response was limited and not sufficient in the cow to fully recover from advance clinical JD. The study besides exhibiting therapeutic potential of goat based ‘indigenous vaccine’, also underlined the critical role of nutrition in response to vaccination and in management of chronic infections, just as it hastens or delays the precipitation of disease from sub-clinical to clinical or advance clinical stage. The study also exposed limitation of laboratory tests used for the diagnosis of clinical BJD.


Mycobacterium avium subspecies paratuberculosis (MAP) cause Johne’s disease (JD) in domestic livestock and other animals including primates (Singh et al., 2013a). The disease is endemic in the population of domestic cattle in the country (Sharma et al., 2008; Mishra et al., 2009; Singh et al., 2013a). Though number of dairy animals is highest in the country and is leading milk producer (117 million tonnes) in the world (FAO, 2012) however, per animal productivity is very low. Majority of dairy cattle are in small herds (3 – 5) or as individual animals in milk. Due to high population density of animals there is drastic reduction in grazing and fallow land. There is marked shift in husbandry practices and animals are confined to either one place or in very limited area, where they are taken daily for grazing or watering. Therefore, in the absence of policy on control of JD and under confined husbandry practices, country is waking up to the new challenge of the chronic infections like JD, since JD has long been considered as the disease of large herds and flocks. In this scenario, JD appears to be major cause of low per animal productivity of the native livestock breeds. This low per animal productivity may be directly correlated with high prevalence of JD in Indian livestock. Low priority to JD is also due to lack of information on incidence of MAP in domestic livestock population of the country. Presently country lacks indigenous, rapid, sensitive and cost-effective diagnostic tests and high cost of imported kits are main reasons for lack of attention on JD. Treatment of disease is both uneconomical and impractical (Harris and Barletta, 2001). Since ‘cow slaughter’ is banned in the country, therefore vaccination is the only alternative for the management of bovine JD infecting cattle. ‘Indigenous vaccine’ developed for the control of disease in goats has also shown promise to treat, manage and control infection of MAP in animals other than goats (Singh et al., 2013b).

In the present case reports, two animals (a young male calf and an adult lactating cow) were vaccinated with ‘indigenous goat based vaccine’ in effort to treat these animals of the clinical bovine Johne’s disease.

An adult dairy cow (14 months old, female) of Hariana breed belonged to a poor and landless farmer of village Begampur, located on the other side of bank of Yamuna river near holy city of Vrindavan in Mathura district of Uttar Pradesh. As per owner this cow had calved >8 times and was still giving 300–500 mL of milk per milking in a day (two times milking/ day) despite suffering from diarrhoea continuously for past three years and has soiled all the wall of the courtyard of his small house. The owner had only this cow as the source of

milk for family and was small time contract farmer to earn livelihood and took cow with him for grazing on farm boundaries. For the treatment of his cow, the farmer was taking help of a local NGO (project Balam), run by CDHIFI (Chandi Duke Hefnerr International Foundation of India) in Vrindavan, Mathura for last 6 months. The cow was treated by veterinary experts of the NGO with antibiotics, antiparasitic (Ivomax injection, Merial, USA), anti diarrhoeal drugs and supportive therapy. However, there was limited response in diarrhoea, till antibiotic tablets lasted. As soon as antibiotics were stopped, there was relapse in diarrhoea intensity. The nutritional status of cow was low as she depended on 8–10 hr grazing (extensive management) and little supplement with wheat straw and some wheat flour, while at home. Physical condition of cow was indistinguishable to that of ‘advance clinical JD’. However, screening of serum, blood and fecal samples of the cow (provided by CDHIFI) twice before vaccination were negative in fecal microscopy and blood PCR but was positive in ‘indigenous ELISA’ test. The cow was in a stage where it looked like ‘run over by the disease’, therefore negative in laboratory tests. On the basis of symptoms and no other option, it was decided to give JD vaccine a try with the consent of owner.

A young male calf of Hariana breed (14 months old) was reared by CDHIFI, the NGO in Vrindavan. Mother of calf died due to dystocia during birth of this calf and was reared on external source of milk. Among livestock the NGO had only this calf. Though the calf was on high plane of nutrition (ad lib quality concentrate ration and fodder and was stall fed) and in good management and hygienic conditions, however, calf was suffering from clinical JD (dark in colour unlike white colour of Hariana breed, physically weak and stunted without diarrhoea). On screening against MAP infection, the calf was negative in fecal microscopy, blood PCR and indigenous ELISA. The calf was regularly given Ivmec as first line of treatment against ecto and endo parasites. The experienced veterinarians of NGO tried all possible therapies for stunted growth, before referring to Animal Health Division of Central Institute for Research on Goats (CIRG), Makhdoom, Dist., Mathura. With the consent of NGO, the calf was vaccinated against JD. ‘Indigenous vaccine’ was developed at CIRG, Makhdoom using native ‘5’ strain, a new biotype (‘Indian Bison type’) of MAP (Sohal et al., 2009; Singh et al., 2013c) for the therapeutic management of JD in goats and sheep (Singh et al., 2010a; Singh et al., 2013b; Shroff et al., 2013). The cow and calf were vaccinated with 2 ml of ‘Indigenous Vaccine’ subcutaneously in middle of neck and were monitored at weekly interval, initially and later at monthly intervals. Monitoring parameters included physical condition, health (status of diarrhoea and weakness), production (growth rate, milk yield), sero-conversion and status of shedding of MAP in feces.

For diagnosis and monitoring following tests were used. In microscopy, nearly 2 gram of fecal sample was finely grounded in sterilized distilled water and centrifuged (4000 rpm for 45 min) at room temperature. Smear was prepared from middle layer, stained by Zielh Neelsen (ZN) method and examined under oil immersion (100X) for acid–fast bacilli (AFB) indistinguishable to MAP. In IS900 blood–PCR, DNA isolated from blood samples was subjected to specific IS900 PCR (Singh et al., 2010b). Presence and yield of specific PCR product (413 bp) was analyzed by 2% agarose ethidium bromide gel electrophoresis. Indirect ELISA was used as per the protocol of Singh et al. (2007b) to know the status of MAP infection (S/P ratio method of Collins, 2002) and monitoring of immune response. S/P ratio of 1.0 to 10.0, 0.4 to 0.99, 0.25 to 0.39 and <0.25, were considered strong positive, positive, low positive and negative, respectively. Animal in positive and strong positive category were considered positive for MAP infection or as sero–converted after vaccination.

The cow was negative in fecal microscopy and blood PCR but was positive in ELISA before vaccination (Table 1). Cow started showing recovery two weeks after vaccination and physical condition improved considerably in two months post vaccination (Fig. 1). Volume of the diarrhoea decreased whereas consistency of diarrhoea increased by 15–20%. Due to floods there was break in monitoring as village was surrounded by water and grazing area was inundated, therefore cow suffered further stress. After 4th month, when we went to see the cow, it has relapsed exhibiting watery diarrhoea and further deterioration in physical condition exhibiting emaciation and hide bound condition (Fig. 3A). The size of ‘take’ was also reduced significantly. In this stage also cow was giving 500 ml of milk twice daily. The deterioration was mainly due to submerging of meager grazing area and it took more than one month for water to recede and still area was marshy and unfit for grazing.

Table 1. Animal profile with respect to MAP infection and post vaccination improvement

<table>
<thead>
<tr>
<th>Parameters</th>
<th>0 Day</th>
<th>30 DPV</th>
<th>60 DPV</th>
<th>120 DPV</th>
<th>180 DPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Status of MAP infection</td>
<td>Health</td>
<td>Diarrhoea (thin, watery &amp; copious) &amp; weak</td>
<td>Diarrhoea reduced &amp; health improved</td>
<td>Intermittent diarrhoea, weakness &amp; emaciation started</td>
<td>Continuous diarrhoea &amp; emaciation</td>
</tr>
<tr>
<td>Status of MAP infection</td>
<td>Microscopy</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Status of MAP infection</td>
<td>Blood PCR</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Status of MAP infection</td>
<td>ELISA</td>
<td>Positive</td>
<td>Negative</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>Status of MAP infection</td>
<td>Blood PCR</td>
<td>NA</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Status of MAP infection</td>
<td>ELISA</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
</tr>
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</table>

Table 1. Animal profile with respect to MAP infection and post vaccination improvement
In advanced stages of disease (clinical and advance clinical) vaccination was not successful (Singh et al., 2010a). In cases optimum nutrition as was case in goats at CIRG, the majority of goats (80–85%) recovered, however 15–20% goats with advance clinical JD were lost and could not recover (Singh et al., 2011).

In the present study also, either initial dose of vaccine was not optimum or cows needed booster dose after initial vaccination or immune system was damaged beyond repair. Since immune system in advance cases of JD is weak and under stress, therefore it needs due consideration before recommending increasing 'initial loading dose' or 'booster dose' as an option. However, in the cow booster vaccination after 2 months did not help, since already stress immune system was further damaged by initial dose. Reduced initial dose might have been a better option. In cow booster vaccine might have further stressed or damaged already fragile immune system leading to hastening the disease process and emaciation. Lactation, milking and

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**Figure 1:** Sequential improvement in the lactating cow after vaccination with JD vaccine. A: Before vaccination; B: 30 Days post vaccination; C: 60 Days post vaccination

**Figure 2:** Sequential improvement in the male calf after vaccination with JD vaccine
depletion of feeding resources were other important stress factors that contributed to the failure of immune response, after the cow showed recovery from diarrhoea and improved in physical condition for two months. Though the cow was ‘over-run’ by disease, yet immune–system was working in initial 2 months after vaccination. Since cow was in positive category for the status of JD by ELISA. Native cow showed greater resilience to sustain the infection despite being in diarrhoea for last three years.

The male calf was negative in all three tests; fecal microscopy, blood PCR and indigenous ELISA (Table 1). This may be due to early stage of infection, one time sampling and dormancy (Singh et al., 2013d). Two weeks after vaccination calf developed very good take and started improving in body condition (Fig. 2). Appetite increased and in next 4 months calf showed extremely good recovery and growth rate (fig. 3B).

Figure 3: Physical appearance and Animal health at different time intervals post vaccination. (A) Farmer’s cow of Begampur, (B) Male calf of CDHIFI, Vrindavan

The calf bloomed in health and became shining white in colour. Monitoring sero-status of calf showed that it sero-converted and peak titers were achieved at 180 days post vaccination. It may be due to stress and partial damage to the immune system (mesenteric lymph nodes) due to MAP infection. Singh et al. (2010a and 2011) has shown that after vaccination all clinically affected goats, sero-converted and achieved peak titers by 75 to 150 days post vaccination using ‘indigenous JD vaccine’. Whereas in prophylactic vaccination of young kids sero-converted and achieved peak titers by 35 days post vaccination (Singh et al., 2007a). In subsequent monitoring calf showed consistent improvement except during July and August when calf could not get proper green fodder due to floods. Even after 7 months of vaccination, calf continued to maintain good ‘tack’ (almost of same size as in the beginning) and good health. Despite calf was negative in laboratory tests but responded to ‘therapeutic management’ of JD using ‘indigenous goat based vaccine’.

The two case reports also exposed the limitation of diagnostic tests in JD and why all the infected animals cannot be detected in one time screening using single test. Therefore, diagnosis on the basis of one or two tests can be misleading and it is always handy to use multiple tests taking physical (weakness, stunting, diarrhoea) and production (low birth weight, low growth rate, lowered productivity) conditions into consideration, is always prudent and sometimes better indicator if used judiciously with discretion. In sub-clinical or initial stages of clinical JD calf though started to show symptoms but was not shedding MAP. Animals on optimum nutrition may not have shedding of MAP in fces or in blood circulation or develop in-sufficient antibody titters. Therefore, it is advisable not to ignore physical symptoms as they are characteristic along with history. Treatment history and follow-up of routine health care schedule are important in diagnosis of JD in individual animals. Weakness in calf without diarrhoea was rare in cattle which is the usual symptom in goats and sheep. Since calf was routinely dewormed, therefore had no predisposing factor for developing diarrhoea. Dose of 2 ml of goat based vaccine with optimum nutrition, seemed to work up to 7 months of vaccination (period for which calf was monitored) (Fig. 3).

Study revealed endemicity of MAP infection in Indian native cattle as reported by Sharma et al. (2008). Study showed that animals suffering with clinical and advance clinical JD can be retrieved back to health and productivity using indigenous vaccine. Inactivated vaccine reduces incidence of clinical JD (Reddadcliff et al., 2006; Singh et al., 2007a; Singh et al., 2013b), is cost effective and has therapeutic potential (Singh et al., 2010a; Bastida and Juste, 2011). Comparative efficacy of ‘indigenous vaccine’ developed at CIRG, Mukhdoom was better over a commercial vaccine for some of parameters analyzed in an experimentally challenged trial in goats (Singh et al., 2007a).

Therefore, indigenous goat based vaccine can be used for therapeutic management of JD in cattle and underlined role of stress factors (principally nutrition) in response to vaccination. Impact of candidate strain of vaccine was also observed by Uzonna et al. (2003) who reported that native field strain based paratuberculosis vaccine was more effective and efficacious than commercial vaccine. On same logic in this study also the goat based indigenous vaccine was used to treat clinical JD.

Study also underlined need for immediate attention for management and control of JD in the population of native domestic cattle. Response to specific treatment (vaccine) in the cow and calf confirmed the initial diagnosis based on clinical symptoms, where specific diagnostic tests failed.

CONFLICT OF INTEREST
No conflict of interest to declare.

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