

Research Article



Comparative Study for Detection of Subclinical Endometritis in Local Cows

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Abstract | The objective of the present study was to determine using the Dramincki estrus detector and to evaluate its efficiency in diagnosis of subclinical endometritis in cows. In addition, the diagnosis of subclinical endometritis with different methods such as assess the efficacy of endometrial cytology to determine the use of percentage of poly morpho-nuclear cells PMNs in cervical mucus, measurement of the PH, White side test and sperm penetration test of the cervical mucus. The study was conducted on 42 cows suffering from regularly repeated estrus located in different regions in Nineveh province from the period of 01/10/2017 to 01/10/2018. All cows were subjected to general clinical examination, cervical and vaginal examination by vaginal speculum, and the Dramincki detector was used to ensure that the cows are in estrum. Samples of cervical mucus were collected from all cows for estimation the percentage of PMNs ($12 \geq 18$). Whiteside test was utilized to determine changes in cervical mucus color from normal to a light yellowish color. Furthermore, microscopic sperm penetration test was used to determine the ability of sperm to penetrate the cervical mucus by adding a drop of liquid semen close to the mucus sample. Results showed that subclinical endometritis was diagnosed in 16.66% (7/42) of the repeated breeding cows. However, there was no significantly correlation between the age of the cows and both number of estrus, and values of the estrus detector in cows affected with subclinical endometritis. Conversely, there was a significant correlation in the detected subclinical endometritis by endometrial cytology that indicated as an increasing percentage of PMNs up to $12 \geq 18\%$ with the increased age of the affected cows at (5.43 ± 0.841). Similarly, subclinical endometritis was highly significant associated with the increased PH value of cervical mucus (7.800 ± 0.0577) as compared with unaffected cows (7.271 ± 0.034), change the mucus color to light yellowish, as well as with the mucus penetration ability of sperms, as the majority of sperms died after a short distance of mucus penetration from cows suffered from subclinical endometritis. To some briefly, diagnosis of subclinical endometritis, using pH value, cytological investigation, white side test and sperm penetration test provided a considerable value in field conditions.

Keywords | Subclinical endometritis, pH, Cytology, Whiteside, Sperm penetration

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INTRODUCTION

Subclinical endometritis considers as one of the important etiological factors of repeat breeding in cows. Early diagnosis is essential to minimize the economic loss. Diagnosis of infectious repeat breeding cows and isolation of the non-infectious repeat breeding problem is first step to challenge for the field veterinarian. Careful checking of uterine ultrasonography scanning of the uterus, vaginos-

copy of vagina, endometrial cytology, uterine mucus and uterine biopsy have been testified as diagnostic approaches to identify endometritis (Barlund et al., 2008). Posing considerable economic loss to dairy farmers. Subclinical endometritis has been the one of major cause the repeat breeding (Arthur et al., 1989).

Repeat breeders are animal cycling normally without any clinical abnormalities, but fail to conceive even after at

least three successive inseminations. They have clinically normal reproductive tract, estrous cycles and estrous periods (Arthur et al, 2001).

Subclinical endometritis alters the physico-chemical properties of cervical mucus and: therefore, examination received of cervical mucus for appearance PH. It maybe valuable in its diagnosis (Vijayajan et al., 2007; Hussain et al., 1991).

Cytological examination of reproductive tract is often used to evaluate possible reproductive lesions by cervical cytological examination in cow (Mateus et al., 2002b, Leblanc et al., 2002a) The endometrial and inflammatory cells detected technique were accepted and controlled by cyto-brush technique and mucus smear (Kamanickam et al., 2005b) techniques to evaluate endometrial cytology specially as an aid in the detection of endometritis prepared smear most contain epithelial cells. A ratio of polymorph nuclear cells (PMNs) to epithelial should be calculated. If the ratio is more than 12% PMNs to epithelial cells, sub-clinical endometritis significantly (Mateus et al., 2002b: Leblanc et al., 2002a).

This clearly indicated the efficacy of white side test WST for detection of subclinical endometritis by boiling point and after cooling the intensity of color changes were graded as normal (no color) mild infection or subclinical endometritis (light yellow color) moderate infection or clinical endometritis (yellow color) severe infection or chronic endometritis (dark yellow color) (Kumar et al., 2015), The color change observed in white side test (WST) in control cows might be due to neutrophil infiltration and metrorrhagia, which are usually seen during estrogen dominance (Ohtani et al., 1993; Raja et al., 2012).

Cervical mucus pH and bacterial load in CM are absolutely correlated with WST and the test is apposite in detection of endometritis at both clinical and subclinical degree. Moreover, White side test can decrease the cost for analyzing endometritis. Detection of endometritis with white side test has clinical importance, and the test can be directed at the field level, when they fail to conceive repeatedly (Raja et al 2012).

IgA-antisperm antibody (ASA) is highly detectable in cervical mucus (CM) (Wang et al., 2009). ASA in cervical mucus has a great impact on the ability of sperms to penetrate cervical mucus in human (Eggert-krust et al., 1991). There are only a little amount of researches in veterinary field under the influence of ASA in CM on sperm penetration and its impact on fertility rates. The principle aims of the present study are to determine the incidence of subclinical endometritis in local cows using various methods for diagnosis. These methods included the using of dramincki

estrus detector, endometrial cytology to determine the percentage of PMNs, measurement the pH, White slide test and sperm penetration test in cervical mucus.

MATERIALS AND METHODS

All repeats breeding cows were observed and data (age cows, number of repeated estrus) was recorded. Forty two regular cyclic local cows were presented to the Teaching Veterinary Clinical Service in College of Veterinary Medicine-Mosul University and Ninawah Veterinary hospital; the study was conducted during the one -year period from October 2017 to October 2018. Gynecological examination of these cows revealed no palpable abnormalities of the reproductive organ.

All cows were subjected to detailed general clinical examination, cervical and vaginal examination by vaginal speculum, and the Dramincki detector was used to ensure that the cows are in estrum. With aseptic precautions, using the sterile AI sheath connected with 20ml syringe, cervical mucus, which represented the endometrial secretion, was collected.

All Mucus samples were estimated of PH value by portable PH meter. Endometrial cytology samples were collected from each cow included in this study aspirated from cervical lumen using sterile catheter and transferred into sterile tubes. Then transported to the laboratory at 4°C. Smears were prepared from the cervical mucus on clean glass microscope slide and fixed with absolute methyl alcohol and stained by Right-Giemsastain (Mateus et al., 2002). Endometrial cytological examination of polymorph nuclear cell (PMNs) was evaluated using light microscope at magnifications 100X and 400X. Whiteside test was used to determine changes in cervical mucus coloration from normal to a light yellowish color. One ml of the cervical mucus was mixed with one ml of 5 per cent sodium hydroxide solution in a test tube and heated up to the boiling point and subsequently cooled in running tap water. The appearance of yellow color was considered as the positive indicator of presence the infection. Depending on the intensity of color development, the degree of endometritis was classified as No color (absence of infection), mild yellow color change (mild infection or subclinical), intense yellow color (severe infection or clinical) (Anilkumar and Devanathan, 1996). Sperm penetration test was used to determine the ability of sperm to penetrate the cervical mucus by adding a drop from straw contain 20×10^6 sperm of liquid semen close to the small amount of mucus sample put on the worm slide at 37° C. and showed the sperm penetrated by microscope with dark field at 100X.

Laboratorial examinations revealed that subclinical endometritis was diagnosed in 7/42 (16.66%) of the repeated breeding local cows. Subclinical endometritis was not diagnosed in cows ranged between 3-8 years old (means 5.00 ± 0.272). Conversely, subclinical endometritis was diagnosed in cows ranged between 3-9 years (means 5.43 ± 0.841). Inseminated unaffected cows were returned to estrus for 2-4 times (means 2.80 ± 0.141) due to conception failure. However, return to estrus, after insemination, was detected 2-5 times (means 3.43 ± 0.369) in the affected cows. In unaffected cows, the estimated values of the Dramincki-estrus detector were ranged over 200-300 (means 254.86 ± 4.326). While, the values in cows affected with subclinical endometritis were 240-250 (means 247.14 ± 1.844). There was no significant correlation between the age of the cows and both numbers of estrus, and values of estrus detector in the unaffected and affected cows. However, PH values of cervical mucus were significantly varied ($P \leq 0.01$) between the unaffected and affected cow, 7.271 ± 0.034 and 7.800 ± 0.0577 , respectively. (Table 1).

Table 1: Shower correlated between Age, Estrus Number, Estrus detection, Endometrial Cytology PMN% and PH- value in unaffected cow compared with Sub-clinical endometritis cow.

Groups Parameters	Unaffected cow	Sub-clinical endometritis cow	P- value
Age\years	5.00 ± 0.272	5.43 ± 0.841	0.084
Estrus number	2.80 ± 0.141	3.43 ± 0.369	0.550
Estrus detection	254.86 ± 4.326	247.14 ± 1.844	0.436
Endometrial Cytology PMNs%	0.20 ± 0.080	$14.86 \pm 1.122^{**}$	0.000
PH- value	7.271 ± 0.034	$7.800 \pm 0.0577^{**}$	0.000

Values are mean \pm S.E. ** mean significant at $P \leq 0.01$

In unaffected cow, the percentage of endometrial PMNs ranged from 0-1% (mean 0.20 ± 0.080); and 12-18% in cows affected with subclinical endometritis. There was a significant correlation between the increased age in the percentage of PMNs up to $12\% \geq 18\%$ in affected cows at (5.43 ± 0.841), forever there was a significant correlation in the detected subclinical endometritis by endometrial cytology indicated as an increasing percentage of PMNs up to $12\% \geq 18\%$ with the increased age of the affected cows at (5.43 ± 0.841). (Table 1, 2) (Figure 1, 2). The White side test was performed in the cervical mucus samples collected at estrus was recorded to change the mucus color to light yellowish, as well as observed in subclinical endometritis light yellow color compared with non-coloration in unaf-

ected cows.

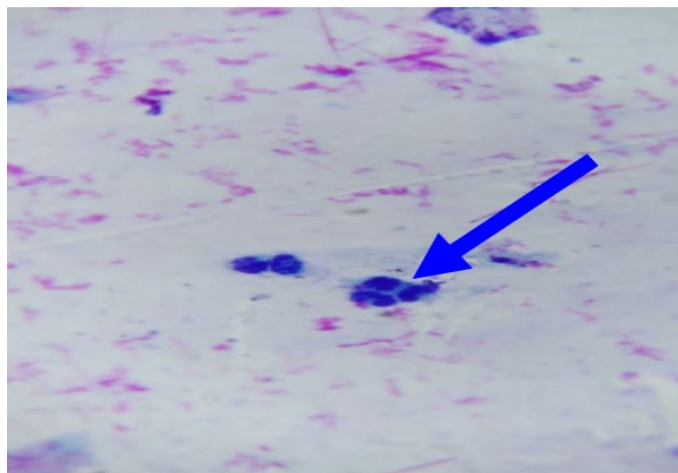


Figure 1: Polymorphonuclear cell (arrow) mucus subclinical endometritis. Giemsa stain X1000

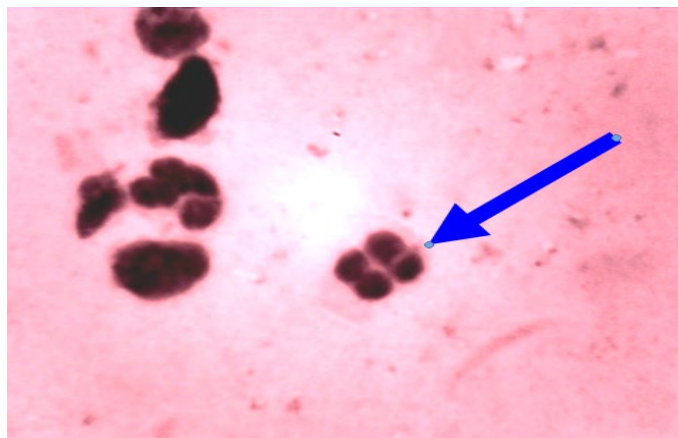


Figure 2: Polymorphonuclear cell (arrow) in cervical mucus subclinical. Right-Giemsa stain X1000

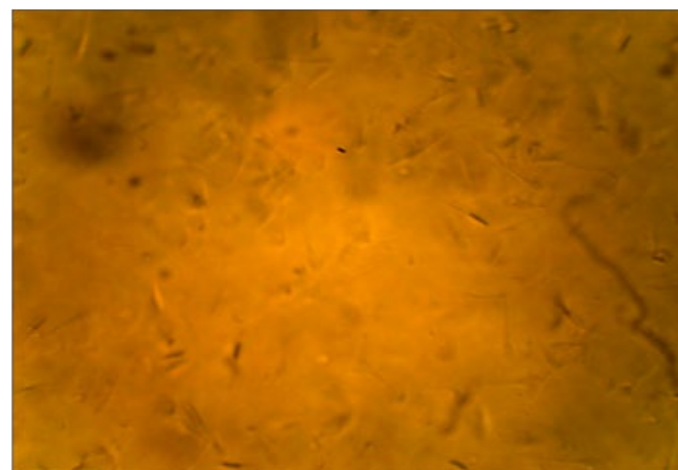


Figure 3: A live sperm penetration majority of cervical mucus in unaffected cow. X400

Penetration was measured using phase contrast microscopy. However the penetration can distinguish samples of frozen-thawed bovine semens were measured at penetrated ability of sperms for clear cervical mucus, whereas sperm

Table 2: Shower correlated between Age, Estrus Number, Estrus detection, Endometrial Cytology PMN%, PH- value White side test and Sperm penetration in unaffected cow compared with Sub-clinical endometritis cow.

Correlations		Groups	Age\years	Estrus number	Estrus detection	Cytological method PMNs%	PH value	White side test
Groups	r - value	1	.270	.095	-.123	.977**	.718**	1.000**
	p - value		.084	.550	.436	.000	.000	.000
Age\years	r - value	.270	1	.494**	-.160	.361*	.374*	.270
	p - value	.084		.001	.312	.019	.015	.084
Estrus number	r - value	.095	.494**	1	-.219	.165	.171	.095
	p - value	.550	.001		.163	.297	.278	.550
Estrus detection	r - value	-.123	-.160	-.219	1	-.126	-.085	-.123
	p - value	.436	.312	.163		.428	.590	.436
Cytological method PMNs%	r - value	.977**	.361*	.165	-.126	1	.745**	.977**
	p - value	.000	.019	.297	.428		.000	.000
PH value	r - value	.718**	.374*	.171	-.085	.745**	1	.718**
	p - value	.000	.015	.278	.590	.000		.000
White side test	r - value	1.000**	.270	.095	-.123	.977**	.718**	1
	p - value	.000	.084	.550	.436	.000	.000	
Sperm penetration	r - value	-1.000**	-.270	-.095	.123	-.977**	-.718**	-1.000**
	p - value	.000	.084	.550	.436	.000	.000	.000

**Correlation is significant at the 0.01 level yellow color.

* Correlation is significant at the 0.05 level orange color.

motility is arrested by subclinical endometritis cervical mucus. It has been found that the majority of sperms died after a short distance of mucus penetration from cows affected by subclinical endometritis while showed the majority of sperms motility were penetrated and survived at the mucus after moderate distance in unaffected cows. (Figure 3,4).

DISCUSSION

A previous study indicated the usefulness of utilizing the changes in the physico-chemical properties of cervical mucus for diagnosis of repeat breeder cow caused by sub-clinical endometritis (Kumar et al., 2015). In this study, the occurrence of subclinical endometritis was 16.66 %, which is consistence (17%) with Madoz et al. (2013). However, higher incidence rates were reported by Dubuc et al. (2010) and Carneiro et al. (2014) (18.7% and 26%, respectively). pH values of cervical mucus were significantly varied ($P \leq 0.01$) between the unaffected and affected cows (7.271 ± 0.034 and 7.800 ± 0.0577 , respectively). Increase of pH in vaginal discharges higher than 7.8 is in agreement with a previous study in cattle affected with endometritis (Singla et al, 2004). This could be due to the presence of bacterial contamination in the uterine fluids where the increased pH is unsuitable for survival of spermatozoa and embryo in the uterus (Sheldon et al., 2006). The increased pH may results in metabolism of bacteria and inflammatory exudates in cervical mucus (Salphale et al., 1993). There was a significant correlation in the detection of subclinical endometritis by endometrial cytology indicated by an increased percentage of PMNs (up to $12\% \geq 18\%$) with the increased age of the affected cows. This factor suggests

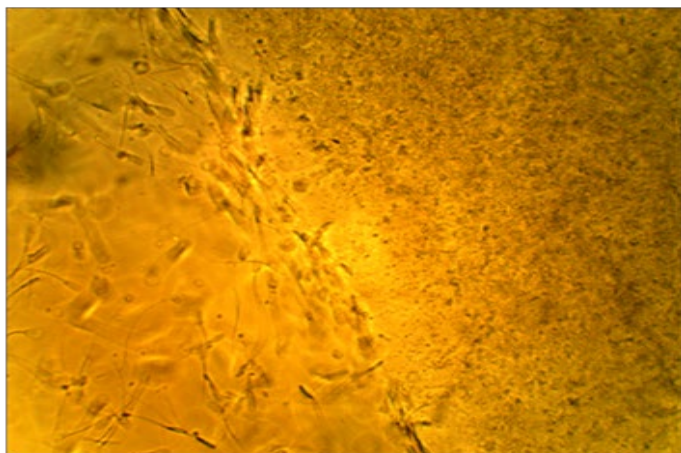


Figure 4: Died sperms after a short distance of mucus penetration in subclinical endometritis cow. X400.

that older cows showed an increased risk of subclinical endometritis. Many studies have stated that endometrial cytology is an effective technique for the initial detection of subclinical endometritis along with the microbial evaluation (Bajaj, 2015). However, the clear mucus discharge of $\geq 10\%$ polymorphonuclear cells (PMNs) are an indicator of subclinical endometritis, (Dutt et al., 2017). Kasimanickam et al. (2004) recorded that $>18\%$ neutrophils count after 20-33 days postpartum, or $>10\%$ neutrophils at 34-47 days postpartum in mucus samples as an indicative of subclinical endometritis; although, Gilbert et al. (2005) establish 5% neutrophils at 40-60 days postpartum as an indicator of subclinical endometritis in cattle. While Barlund et al. (2008) used a neutrophil threshold value of 8% at 28-41 days postpartum in cattle to declare endometritis.

All the affected cows in this study were positive (100%) for white side test, indicating positive for subclinical endometritis, where the coloration of the cervical mucus was changed to a light yellowish color, as compared with non-coloration in unaffected cows. This obviously indicated the ability of white side test in detection of subclinical endometritis, (Raja et al., 2012). Positive reaction to white side test might be due to neutrophil infiltration and metrorrhagia, which are usually seen during estrogen dominance (Ohtani et al., 1993). The normal cervical mucus contains fewer numbers of leukocytes to cause such changes in color, although in clinical and subclinical endometritis, cervical mucus contains a high number of leukocytes producing a color reaction (Pateria and Rawal, 1990). The sperm penetration test of frozen-thawed bovine semen was measured at penetration ability of sperms in clear cervical mucus, whereas sperm motility is arrested by subclinical endometritis cervical mucus on the worm slide at 37°C . It has been found that the majority of sperms died after a short distance of mucus penetration collected from cows affected with subclinical endometritis. In contrast, in unaffected cows, the majority of the penetrated sperms were survived after moderate distance in the mucus. Cellular accumulation and leukocytes might limit the migration of spermatozoa in cervical mucus (Muzaffer, 2007). Therefore, it was recommended to utilize this test to distinguish between mucus collected from suspected subclinical endometritis in cows (Anilkumar and Devanathan, 1996). Presence of IgA-anti sperm antibodies can inhibit passage of spermatozoa through cervical mucus, prevent membrane fluidity changes needed for capacitation, and reduce the ability of spermatozoa to undergo the acrosome reaction, (Fijak and Meinhardt, 2006).

CONCLUSION

This study evaluated the using of different techniques for diagnosis of subclinical endometritis. In particular, for clinical practice, sperm penetration test was recognized as

the most practical and simple method for detection of subclinical endometritis.

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CONFLICT OF INTEREST

Authors declare no conflict of interest.

AUTHORS CONTRIBUTION

The authors themselves took the design of the study and collected the data and examined the cases and analyzed statistically and research writing

REFERENCES

- Anilkumar R, Devanathan TG (1996). Correlation between White Side test for quality of cervical mucus and sperm penetration test. *Indian Vet. J.* 73: 1099-1100.
- Arjunraot TG (2017). Efficacy of Neem extract, logul's iodine and *E.coli* LPS for the treatment of endometritis in cows and buffaloes. Master of Veterinary Science in Animal Reproduction, Gynecology and Obstetrics. College of Veterinary and Animal Sciences, Parbhani-431 402.
- Arthur GH, Noakes DE, Pearson H (1989). *Veterinary Reproduction and Obstetrics*. 6th ed. Billiere, Tindall, London. pp. 384-388.
- Arthur GH, Noakes DE, Pearson H (2001). *Veterinary Reproduction and Obstetrics*. 8th edition, Bailliere Tindall, London, Pp. 212.
- Bajaj NK (2015). Diagnosis and therapeutic management of sub-clinical endometritis in postpartum buffaloes. Ph.D. (Veterinary Gynaecology and Obstetrics), Nanaji Deshmukh Veterinary Science University, Jabalpur (M.P.).
- Barlund CS, Carruthers TD, Waldner CL, Palmer CW (2008). A comparison of diagnostic techniques for postpartum endometritis in dairy cattle. *Theriogenology*. 69: 714-723. <https://doi.org/10.1016/j.theriogenology.2007.12.005>
- Barlund CS, Carruthers TD, Waldner CL, Palmer CW (2008). A comparison of diagnostic techniques for postpartum endometritis in dairy cattle. *Theriogenology*. 69:714-723. <https://doi.org/10.1016/j.theriogenology.2007.12.005>
- Carneiro LC, Ferreira AF, Padua M, Saut JPM, Ferraudo AS, dos Santos RM (2014). Incidence of subclinical endometritis and its effects on reproductive performance of crossbred dairy cows. *Trop. Anim. Health Prod.* 46:1435-1439. <https://doi.org/10.1007/s11250-014-0661-y>
- Dubuc J, Duffield TF, Leslie KE, Walton JS, Leblanc SJ (2010). Risk factors for postpartum uterine diseases in dairy cows. *J. Dairy Sci.* 93: 2764-5771. <https://doi.org/10.3168/jds.2010-3429>
- Dutt R, Singh G, Singh M, Sharma M, Dalal J, Chandolia

- RK (2017). Diagnosis of Subclinical Endometritis in Murrah Buffaloes through Cytobrush Technique. *Int. J. Curr. Microbiol. App. Sci.* 6(11): 494-49. <https://doi.org/10.20546/ijcmas.2017.611.059>
- Eggert-Kruse W, Hofsäss A, Haury E, Tilgen W, Gerhard I, Runnebaum B (1991). Relationship between local antisperm antibodies and sperm mucus interaction *in vitro* and *in vivo*. *Hum. Reprod.* 6(2): 267-276. <https://doi.org/10.1093/oxfordjournals.humrep.a137320>
 - Fijak M, Meinhardt A (2006). The testis immune privilege. *Immunol. Rev.* 213: 166-173. <https://doi.org/10.1111/j.1600-065X.2006.00438.x>
 - Gilbert RO, Shin ST, Guard CL, Erb HN, Frajblat M (2005). Prevalence of endometritis and its effects on reproductive performance of dairy cattle. *Theriogenology.* 64: 1879-1888. <https://doi.org/10.1016/j.theriogenology.2005.04.022>
 - Hussain AM, Daniel RCW (1991). Bovine endometritis current and future alternative therapy. *J. Vet. Med. Ser. 3:* 641-651.
 - Kasimanickam R, Duffield TF, Foster RA, Gartely CJ, Leslie KE, Walton JS, Johnson WH (2004). Endometrial cytology and ultrasonography for detection of subclinical endometritis in postpartum dairy cattle. *Theriogenology.* 62: 9-23. <https://doi.org/10.1016/j.theriogenology.2003.03.001>
 - Kasimanickam R, Duffield TF, Foster RA, Gartely CJ, Leslie KE, Walton JS, Johnson WH, (2005). A comparison of the cytobrush and uterine lavage techniques to evaluate endometrial cytology in clinically normal postpartum dairy cows. *Can. Vet. J.* 46: 255-259.
 - Kumar S, Bhardwaj A, Srivastava AK, Rao M, Kumar N (2015). Whiteside test: a field test on the cervical mucus of cows for diagnosis of endometritis. *Intas Polivet.* 16(2): 207-213.
 - LeBlanc SJ, Duffield TF, Leslie KE, Bateman KG, Keefe GP, Walton WH, Johnson WH (2002). The effect of treatment of clinical endometritis on reproductive performance in dairy cows. *J. Dairy Sci.* 85: 2237-2244. [https://doi.org/10.3168/jds.S0022-0302\(02\)74303-8](https://doi.org/10.3168/jds.S0022-0302(02)74303-8)
 - Madoz LV, Uliodori GI, Jaureguiberry MJ, Plöntzke M, Drillich J, Sota RL (2013). The relationship between endometrial cytology during estrous cycle and cutoff points for the diagnosis of subclinical endometritis in grazing dairy cows. *J. Dairy Sci.* 96: 4333-4339. <https://doi.org/10.3168/jds.2012-6269>
 - Muzaffer T, Suleyman B, Umüt Cirit, Ozen Banu Ozdas, Kemal Ak (2007). Relationship between bovine fertility and the number of spermatozoa penetrating the cervical mucus within straws. *Anim. Reprod. Sci.* 101:18-27. <https://doi.org/10.1016/j.anireprosci.2006.08.020>
 - Mateus L, Lopes da Costa L, Carval H, Serra P, Robolo Silva J (2002). Blood and intrauterine leukocyte profile and function in dairy cows that spontaneously recovered from postpartum endometritis. *Reprod. Dom. Anim.* 37: 176-181 <https://doi.org/10.1046/j.1439-0531.2002.00351.x>.
 - Ohtani S, Okuda K, Nishimura K, Mohri S (1993). Histological changes in bovine endometrium during the estrous cycle. *Theriogenology.* 39: 1033-1042. [https://doi.org/10.1016/0093-691X\(93\)90004-O](https://doi.org/10.1016/0093-691X(93)90004-O)
 - Pateria AK, Rawal CVS (1990). White side test for subclinical mastitis in buffaloes. *Indian J. Anim. Reprod.* 11: 142-144.
 - Raja S, Devanathan TG, Kulasekar K, Pazhanivel N, Balachandran C (2012). Whiteside Test and Endometrial biopsy for diagnosis of endometritis in repeat breeding cows. *Indian J. Anim. Reprod.* 33(1): 56-58.
 - Salphale GB, Kadu MM, Fasihddin M, Kadu MS (1993). Study of some physical properties of estrous cervical mucus in synchronized animals and Repeat breeder cross bred cows with reference to fertility. *Indian J. Anim. Reprod.* 14: 77-78.
 - Sheldon IM, Lewis GS, Le Blanc S, Gilbert RO (2006). Defining Post-Partum Uterine Diseases in Cattle. *Theriogenology.* 65: 1516-1530. <https://doi.org/10.1016/j.theriogenology.2005.08.021>
 - Singla P, Singh Jagir, Sharma NS, Dhaliwal GS, Kumar Ajeet (2004). Effect of Post A.I. Immunotherapy on Dynamics of Uterine Microflora and Conception in Subclinical Endometritis Cows. *Indian J. Anim. Sci.* 74: 706-770.
 - Vijayarajan A, Chandrhasan C, Ezakial Napoleon RZ (2007). Effect of pre- and post-insemination substitution of GnRH in repeat breeding buffaloes. *Indian Vet. J.* 84: 940-943.
 - Wang M, Shi JL, Cheng GY, Hu YQ, Xu C (2009). The antibody against a nuclear autoantigenic sperm protein can result in reproductive failure. *Asian J. Androl.* 11(2): 183-192. <https://doi.org/10.1038/aja.2008.59>